

## Kinetic Spectrophotometric Determination of Trace Amounts of Sulfide

Mohsen Barzegar,\* Ali Jabbari,<sup>†</sup> and Majid Esmaeili<sup>‡</sup>

Department of Food Science and Technology, Tarbiat Modarres University, P.O. Box 14115-336, Tehran, Iran

<sup>†</sup>Department of Chemistry, K. N. Toosi University, Tehran, Iran

<sup>‡</sup>Department of Chemistry, Razi University, Kermanshah 67149, Iran

Received February 25, 2003

A method for the determination of trace amount of sulfide based on the addition reaction of sulfide with methyl green at pH 7.5 and 25 °C is described. The reaction is monitored spectrophotometrically by measuring the decrease in absorbance of the dyestuff at 637 nm by the initial rate and fixed time method. The calibration graph is linear in the range 30-1200 ppb. The theoretical limit of detection was 0.014 ppm. Seven replicate analysis of a sample solution containing 0.70 ppm sulfide gave a relative standard deviation of 1.5%. The interfering effects of various ions on sulfide determination have been reported and procedures for removal of interference have been described. The proposed method was applied successfully to the determination of sulfide in tap and wastewater samples.

**Key Words :** Sulfide, Kinetic spectrophotometric, Methyl green, Slope method, Fixed time method

### Introduction

Hydrogen sulfide is a by-product of many industrial processes and it is also formed during putrefaction of proteinaceous substances containing sulfur.<sup>1</sup> This hazardous gas can also be produced when elementary sulfur or sulfur containing compounds react with organic material at high temperature. The monitoring of sulfide is required in a variety of environmental and industrial applications, including odor assessments, health and safety investigations, routine industrial/off-line monitoring programs, and regulatory compliance issues as a consequence of the toxicity of any liberated hydrogen sulfide.<sup>2</sup> There are limits on the total level of sulfide permitted in waste discharges in most countries because of toxicity and capacity to remove dissolved oxygen.<sup>3,4</sup> Sulfide in H<sub>2</sub>S form is toxic to fish and other aquatic organisms<sup>5</sup> and can be responsible of the decrease in wastewater treatment plants efficiency. Hence, its concentration needs to be controlled, especially in water and wastewater.

Sulfide can be determined in different media, using various techniques. These include titrimetric,<sup>6</sup> flow-injection analysis,<sup>7,8</sup> amperometric,<sup>9</sup> polarographic,<sup>10</sup> ion chromatography,<sup>11,12</sup> HPLC,<sup>13</sup> GC<sup>14</sup> and spectrophotometric.<sup>15,16</sup>

Although chromatographic methods have low detection limits, they generally require complex and time-consuming sample preparation procedures, are expensive and are not easily adapted to field analysis.

Kinetic methods have been widely used in catalytic and non catalytic determination of various chemical species.<sup>17-19</sup> The different techniques and procedures associated with this type of method have been reviewed.<sup>20,21</sup> Kinetic methods for the determinations of micro amounts of sulfide have been described.<sup>22-24</sup> However, most of these methods suffer from

interfering effects of other sulfur anions or low linear dynamic range and some are time-consuming. Thus, simple, rapid, and selective methods are still required.

In this work we report a kinetic spectrophotometric method based on the addition reaction of sulfide with methyl green at pH 7.5 for the determination of sulfide in trace levels. Application of the developed method was evaluated by measuring the sulfide content in tap water and wastewater samples.

### Experimental Section

**Materials.** All reagents were analytical-reagent grade (from Merck or Aldrich) and used without further purification. Doubly distilled water was used throughout the experiment.

Sulfide stock solution, (1000 ppm) was prepared daily by dissolving 0.7506 g of Na<sub>2</sub>S·9H<sub>2</sub>O (Merck) in water and diluting to 100 mL. This solution can be standardized iodometrically<sup>25</sup> or potentiometrically.<sup>26</sup> Standards of working solution were made by appropriate dilution daily as required.

Methyl green stock solution (Merck, 1.0 × 10<sup>-4</sup> M) was prepared by dissolving 0.0608 g of methyl green in water and diluting to 100 mL in a calibrated volumetric flask. Ten mL of this solution was diluted to 100 mL and used as a working solution. Phosphate buffer (pH 7.5) was prepared by mixing a suitable amount of K<sub>2</sub>HPO<sub>4</sub> (0.01 M) and KH<sub>2</sub>PO<sub>4</sub> (0.01 M) in a 100 mL volumetric flask.

**Apparatus.** The absorption spectra were recorded on a Cecil 9000 UV-Vis double beam spectrophotometer (Cecil Instruments Limited, England) and measurements at a fixed wavelength were made with a 662 Metrohm probe type photometer. The Metrohm photometer was connected to a computer using a 12 Bit A/D card (model 1201, Bitpardaz Co., Tehran, Iran) and the appropriate software was written in Quik Basic in our laboratory. The changes in absorbance

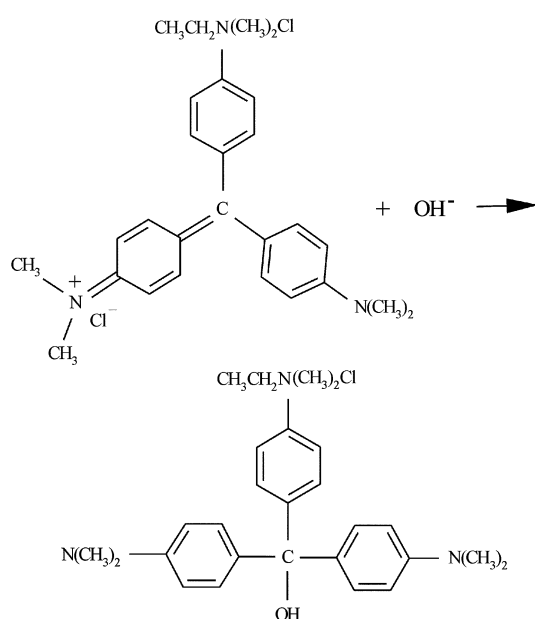
\*Corresponding author. Fax: +98 21 4196524; e-mail: mbb@modares.ac.ir

with time were displayed on the screen. A magnetic stirrer (model ZMS 74, ZAG CHIMIE Co., Tehran, Iran) was used for mixing of reagents and pH was adjusted using a 691 Metrohm pH meter. The temperature was controlled by a thermostated water bath (model HAAKE D1). The temperature was maintained constant in the reaction cell by circulating water at appropriate temperature around the cell of the photometer during the experiment. The addition of sulfide solution to reaction cell was carried out using Hamilton syringes with different volumes.

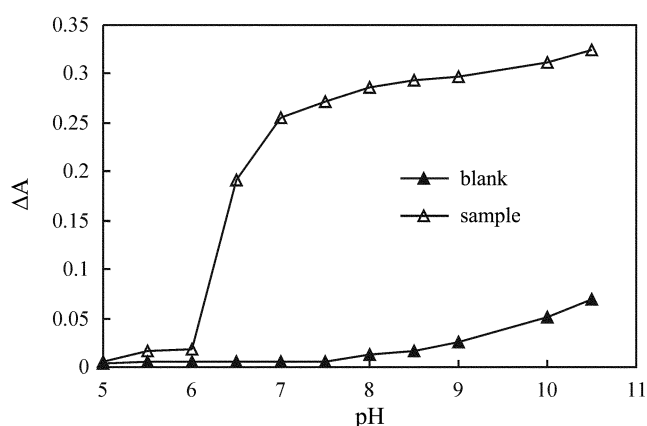
**Procedure.** Two separate procedures were adopted. Both of two procedures were performed by direct data collection by computer. The first method was based on the conventional fixed time method and the second on the slope method. In the cell of reaction, 2.0 mL ( $1.0 \times 10^{-5}$  M) of methyl green solution was added to 1.0 mL of 0.01 M phosphate buffer (pH 7.5) and diluted to 7.0 mL by addition of appropriate amounts of water. Probe was inserted in the cell and the deviation in absorbance of blank was saved in computer. A magnetic stirrer was used to mix the reagents. Then 50-100  $\mu$ L of sulfide solution was added to initiate the reaction and the absorbance versus time data were collected by the computer. With a fixed time method, absorbance changes were measured between 0-15 s after start of the reaction. In the slope method, the slope of the plot of absorbance versus time was found during the first 10 s after starting the reaction.

## Results and Discussion

Methyl green is an acid-base indicator and exists in two



forms,  $R^+$  and ROH, depending on the pH of solution.<sup>27</sup> Addition of sulfide to methyl green occurs at pH 6.0-8.0 (at 25 °C). This causes a relatively rapid change of the color of solution from blue to colorless even in the presence of small



**Figure 1.** Effect of pH on the reaction rate. Conditions: [methyl green]  $3.3 \times 10^{-6}$  M; 0.50 ppm sulfide;  $T = 25$  °C; fixed time 15 s.

amounts of sulfide (ppb).

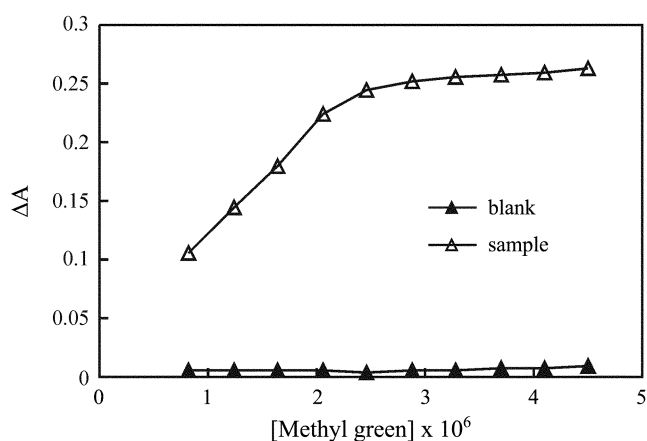
**Effect of Variables.** The effect of pH, concentration of methyl green, ionic strength and temperature on the additional reaction of sulfide with methyl green was studied by the fixed time and slope method.

The decrease of absorbance of methyl green in the presence and absence of sulfide was measured in the pH range 5.0-10.0 using phosphate buffer, HCl, or NaOH solutions for pH adjustments. The effect of pH on the rate of reaction of  $3.3 \times 10^{-6}$  M methyl green in the presence of 0.50 ppm sulfide is shown in Figure 1. As it is seen, the difference between blank and sample reaction rate was increased with increasing pH from 5.0 through 7.5. So pH 7.5 was used as optimum pH because of maximum difference between the rate of blank and sample reaction. Above pH 7.5 the rate of blank reaction is increased owing to the formation of ROH.

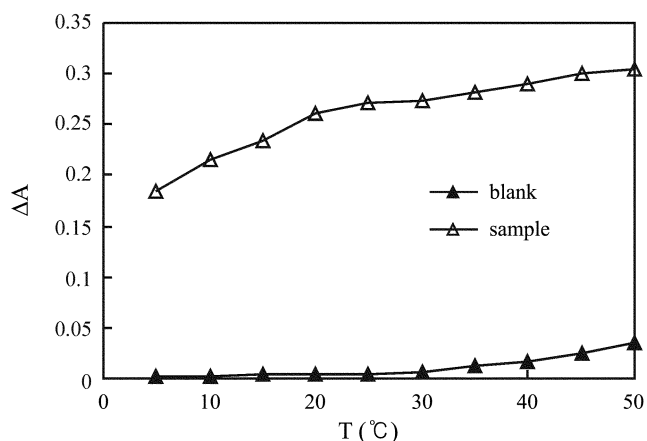
The influence of methyl green concentration on the reaction rate at pH 7.5 was also investigated and the results are shown in Figure 2. It is seen that there was an increase in the reaction rate when the concentration of methyl green was varied from  $0.8 \times 10^{-6}$ – $3.3 \times 10^{-6}$  M while at higher concentration of methyl green no considerable change in the reaction rate was observed. Thus the optimum concentration of methyl green was chosen as  $3.3 \times 10^{-6}$  M.

The effect of ionic strength on the rate of reaction rate was studied. The ionic strength was varied from 0.01-0.10 M using NaNO<sub>3</sub> solution. The results showed that this factor had no effect on the reaction rate up to 0.04 M. A further increase of ionic strength caused a decrease in the reaction rate.

The effect of temperature on the reaction rate was investigated in the range 5-50 °C. Figure 3 shows that the reaction rate of sulfide increased with increasing temperature while the blank had no considerable change up to 25 °C. Temperature of 25 °C was selected as the best working temperature, since at this temperature the differences between blank and sample reaction rate is large and it is suitable for subsequent uses. Optimum conditions were chosen as follows: pH 7.5; [methyl green] =  $3.3 \times 10^{-6}$  M; temperature 25 °C; measuring time 15.0 and 10.0 s for fixed time and slope method,



**Figure 2.** Effect of methyl green concentration on the reaction rate. Conditions: pH 7.5; 0.50 ppm sulfide; T = 25 °C; fixed time 15 s.



**Figure 3.** Influence of temperature on the rate of reaction. Conditions: pH 7.5; 0.50 ppm sulfide; [methyl green] =  $3.3 \times 10^{-6}$  M; fixed time 15 s.

respectively;  $\lambda_{\max} = 637$  nm.

#### Calibration Graph, Precision, and Limit of Detection.

Under optimum conditions two linear calibration graphs were prepared (Table 1). Seven replicate analysis of a sample solution containing 0.70 ppm sulfide gave a relative standard deviation of 1.5%. The theoretical limit of detections<sup>28</sup> were 0.014 and 0.017 ppm for fixed time and slope method, respectively according to  $Y_{DL} = Y_B + E_B$  where  $Y_{DL}$ ,  $Y_B$ , and  $E_B$  are signal of detection limit, blank signal and noise of the blank, respectively. As the results imply, the fixed time method and slope methods are comparable with each other.

**Effect of Foreign Ions.** The interference due to several cations and anions was studied in detail. Different amounts of ionic species were added to the sulfide solution. The concentration of sulfide was 0.50 ppm in solution. The tolerance limit was taken, as the amount required causing  $\pm 4\%$  error in the sulfide recovery. The results are presented in the Table 2.

**Removal of Interference's.** The most of the cations that interfere in sulfide determination form insoluble metal sulfides. Interference effects of these cations were removed

**Table 1.** Analytical characteristics of the elaborated method

	Calibration graph (fixed time method)	Calibration graph (slope method)
Equation	$\Delta A = 1.03 \times 10^{-3} + 5.43 \times 10^{-4} C^a$	$dA/dt = -4.81 \times 10^{-4} + 4.93 \times 10^{-5} C^a$
Linear range (ppb)	30-1200	40-1200
$N^b$	16	16
Correlation coeff.	0.9987	0.9985
RSD%	0.9-2.3 <sup>c</sup>	0.85-2.2 <sup>c</sup>
LOD (ppb)	14.0	17.0

<sup>a</sup>Concentration of sulfide in ppb. <sup>b</sup>Number of calibration points. <sup>c</sup>For sulfide concentration of 50-1100 ppb.

**Table 2.** Effect of foreign ions on the reaction rate of 0.50 ppm sulfide

Foreign ion	Tolerance limit (ppm)
$Na^+$ , $K^+$ , $Li^+$ , $CS^+$ , $Cl^-$ , $F^-$ , $I^-$ , $Br^-$	1000 <sup>a</sup>
$HCO_3^-$ , $S_2O_3^{2-}$ , $SO_4^{2-}$ , $CH_3COO^-$ , $NO_3^-$ , $SCN^-$ , $NO_2^-$ , W (VI), La (III)	800 <sup>a</sup>
$B_4O_7^{2-}$ , $CN^-$ , $PO_4^{3-}$ , $HPO_4^{2-}$ , $ClO_4^-$ , $C_2O_4^{2-}$ , tartrate, $NH_4^+$ , $Mg^{2+}$	500
EDTA, Mo (VI), Cr (VI)	200
Formaldehyde	100
$Sr^{2+}$ , $Ni^{2+}$ , $Fe^{2+}$ , Zr (IV)	50
$Al^{3+}$	10
$Cu^{2+}$	5
$SO_3^{2-}$ , $IO_3^-$	Interfered
$Pb^{2+}$ , $Hg^{2+}$ , $Ag^+$ , $Hg^+$	Interfered

<sup>a</sup>Maximum concentration studied.

by the addition of iodide or hydroxide ions and filtering the solution prior to each measurement. These cations also can be successfully removed from the solution by passing it from a column containing a strongly acidic cation exchanger of  $H^+$  form (Merck). Among the anions, sulfite and iodate interfere. Sulfite interference can be eliminated by the addition of 0.4 mL of 0.05 % formaldehyde to 7.0 mL of test solution prior to each measurement. The interfering effect of iodate was removed by the addition of 1.0 mL 500 ppm iodide to 7.0 mL of test solution prior to each measurement. Since cyanide did not interfere, the interfering effect of silver was removed by the addition of cyanide to form stable silver cyanide complex and thus preventing silver sulfide formation. In addition, the interfering effects of  $Pb^{2+}$ ,  $Cu^{2+}$  and  $Hg^+$ ,  $Hg^{2+}$  were removed by the addition of hydroxide (100 ppm) and iodide (100 ppm), respectively, to form precipitates and thus preventing metal sulfide precipitation. The results are summarized in Table 3.

**Application.** To confirm the usefulness of the suggested method, sulfides were determined in tap water and wastewater samples. After proper dilution of the sample, the wastewater sample was first treated with NaOH and filtered. The pH of filtrate was then adjusted to about 7.5 with HCl. Results are given in Table 4. The results show that the method is suitable for the analysis of real samples.

**Table 3.** The accuracy after removal of interfering ion (0.50 ppm sulfide)

Interfering ion	Masking agent	Sulfide found (ppm)	RSD% (n = 3)
SO <sub>3</sub> <sup>2-</sup> (2) <sup>a</sup>	Formaldehyde (0.003%) <sup>a</sup>	0.486	1.20
IO <sub>3</sub> <sup>-</sup> (5)	I <sup>-</sup> (100)	0.475	1.10
Ag <sup>+</sup> (10)	CN <sup>-</sup> (50)	0.485	1.60
Pb <sup>2+</sup> (10)	OH <sup>-</sup> (100)	0.502	0.95
Hg <sup>+</sup> (10)	I <sup>-</sup> (100)	0.493	1.14
Cu <sup>2+</sup> (10)	OH <sup>-</sup> (100)	0.495	2.10
Hg <sup>2+</sup> (10)	I <sup>-</sup> (100)	0.490	1.60

<sup>a</sup>The term in parenthesis are concentration of interfering or masking agents in ppm except for formaldehyde.

**Table 4.** Analysis of real samples by developed method<sup>a</sup>

Sample	Proposed method		Standard method <sup>b</sup>
	Fixed time	Slope	
Tap water	18.73	18.26	18.53
Wastewater	31.60	32.64	32.02

<sup>a</sup>Five replicate determinations. <sup>b</sup>Standard method [25, 28].

### Conclusions

This work describes a relatively inexpensive, rapid, simple, and sensitive method for the determination of trace amounts of sulfide based on the addition reaction with methyl green. The analytical system is simple and needs minimum maintenance and most of ions do not interfere.

### References

- Puacz, W.; Szahun, W.; Linke, K. *Analyst* **1995**, *120*, 939.
- Patnaik, P. A. *Comprehensive Guide to the Hazardous Properties of Chemical Substances*, 2nd ed.; Wiley: New York, 1999.
- Ebdon, L.; Hill, S. J.; Jameel, M.; Corns, W. T.; Stockwell, P. B. *Analyst* **1997**, *122*, 689.
- Kirk, R. E.; Othmer, D. F. *Encyclopedia of Chemical Technology*, 3rd ed.; Wiley: New York, 1981; vol. 17.
- Eaton, A. D.; Clesceri, L. S.; Greenberg, A. E. *Standard Methods for the Examination of Water and Wastewater*, 19nd ed.; American Public Health Association: Washington, 1995.
- Liu, C. H.; Shen, S. *Anal. Chem.* **1964**, *36*, 1652.
- Lambert, J. L.; Manzo, D. T. *Anal. Chim. Acta* **1969**, *48*, 185.
- Roman Ceba, M.; Vinagre Java, F.; Munoz Leyva, J. A. *Analyst* **1982**, *107*, 781.
- Kurzawa, J. *Anal. Chim. Acta* **1985**, *173*, 343.
- Canterford, D. R. *Anal. Chem.* **1975**, *47*, 88.
- CaSELLA, I. G.; Guascito, M. R.; Desimoni, E. *Anal. Chim. Acta* **2000**, *409*, 27.
- Steinmann, P.; Shoty, W. J. *Chromatogr. A* **1995**, *706*, 287.
- Tang, D.; Santschi, P. D. *J. Chromatogr. A* **2000**, *883*, 305.
- Radford-Knoery, J.; Cutter, G. A. *Anal. Chem.* **1993**, *65*, 976.
- Wood, C. F.; Marr, I. L. *Analyst* **1988**, *113*, 1635.
- Koh, T.; Miura, Y.; Yamamuro, N.; Takaki, T. *Analyst* **1990**, *115*, 1133.
- Barzegar, M.; Rahmani, A.; Jabbari, A.; Mousavi, M. F. *Pharmazie* **2003**, *58*, 114.
- Jabbari, A.; Barzegar, M.; Rahmani, A.; Mousavi, M. F. *Indian J. Chem.* **2002**, *33A*, 2303.
- Barzegar, M.; Mousavi, M. F.; Nemati, A. *Microchem. J.* **2000**, *65*, 159.
- Bendito, D. P.; Silva, M. *Kinetic Methods in Analytical Chemistry*; Horwood: Chichesters, 1988.
- Yatsimirski, K. B. *Kinetic Methods of Analysis*; Pergamon: Oxford, 1966.
- Han, K.; Koch, W. F. *Anal. Chem.* **1987**, *59*, 1016.
- Lei, W.; Dasgupta, P. K. *Anal. Chim. Acta* **1989**, *226*, 165.
- Ensafi, A. A. *Anal. Lett.* **1992**, *25*, 1525.
- Jeffery, G. H.; Bassett, J.; Mendham, J.; Denney, R. C. *Vogel's Textbook of Quantitative Chemical Analysis*, 5nd ed.; John Wiley & Sons: New York, 1989; p 398.
- Hseu, T. M.; Rechnitz, G. A. *Anal. Chem.* **1968**, *40*, 1055.
- Cheng, K. L.; Uendo, K.; Imamura, T. *Handbook of Organic Analytical Reagents*; CRC Press Inc.: 1982; p 462.
- Miller, J. N.; Miller, J. C. *Statistics and Chemometrics for Analytical Chemistry*; Pearson Education Limited: England, 2000.