## Two New Lignans from the Roots of Saururus chinensis

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*Saururus chinensis* (Saururaceae) is a perennial herbaceous plant that has been used in the treatment of various diseases such as edema, jaundice, gonorrhea, fever, and inflammation in Korean folk medicine.<sup>1</sup> Chemical studies of the genus *Saururus* have shown the presence of lignans,<sup>2-6</sup> aristolactams, flavonoids, anthraquinones, and fruanoditerpenes,<sup>7-10</sup> some of which exhibited neuroleptic,<sup>11</sup> hepatoprotective,<sup>12</sup> antifeedant<sup>13</sup> and antioxidant activities.<sup>14</sup> Previously, we reported the isolation of protective agents against sepsis in the animal model from this plant.<sup>15</sup>

Two lignans (1 and 2) were isolated from the EtOAc fraction of the roots of S. chinensis by repetitive column chromatography. Compound 1 was obtained as yellow oil. It showed the value of  $[a]_{D}^{20}$  -65.7° (*c* 1.806, CHCl<sub>3</sub>). A molecular formula of 1 was found to be  $C_{20}H_{22}O_5$  by HRFABMS (*m/z*; found 343.1538 [M+H]<sup>+</sup>; calcd. 343.1545). The UV spectra of 1 revealed the presence of phenolic groups (234 and 287 nm). The <sup>1</sup>H-NMR spectrum showed the presence of two methyl doublets (H-9 and H-9'), two methine groups (H-8' and H-8), one benzylic methylene (H-7'), one benzylic methine group substituted by oxygen (H-7) and two 3,4-methylenedioxyphenyl groups. The 2D <sup>1</sup>H-<sup>1</sup>H COSY spectrum indicated coupling between the oxygenated methine at  $\delta$  4.25 (H-7) and the methine signal at  $\delta$  1.75 (H-8), which was also coupled to the methyl signal at  $\delta$  0.54 (H-9). On the other hand, the methine signal at  $\delta$  2.39 (H-8') was coupled to the methyl signal at  $\delta 0.81$  (H-9'). These observations are consistent with a lignan-7-olic skeleton. The positions of each functional group were determined by a HMBC experiment. One set of correlations was observed between the oxygenated methine H-7 ( $\delta$  4.25) and C-1, C-2, C-6, C-8, C-8' and C-9 and the other set was between H-7' ( $\delta$  2.46) and C-1', C-2', C-6', C-8, C-8' and C-9'. Additionally, the correlations between two methylenedioxy signal at  $\delta$  5.88 and 5.89 and C-3'/C-3 and C-4'/C-4 supported the proposed link the 3,4,3',4'-dimethylenedioxy phenyl moiety. The absolute stereochemistry at C-7

 Table 1. Characteristic <sup>1</sup>H-NMR data of Mosher esters of 1 for determination of stereochemistry

Position	7	9	7′	8′	9′
$1_{S}(\delta_{S})$	5.58	0.53	2.32	1.77	0.68
$1_{R}$ ( $\delta_{S}$ )	5.52	0.57	2.42	2.09	0.78
$\Delta\delta \ (\delta_S - \delta_R)$	S	-0.04	-0.10	-0.32	-0.10

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of **1** was established by modified Mosher's method.<sup>16-18</sup> The differences of chemical shift values obtained by subtracting (*R*)-MPTA ester from (*S*)-MTPA ester  $[\Delta \delta_{\rm H} (\delta_{\rm S} - \delta_{\rm R})]$  are shown in Table 1, and the negative values of  $\Delta \delta_{\rm H} (\delta_s - \delta_R)$  at H-9, H-7', H-8' and H-9' suggested a S configuration at C-7 in compound 1. To determine the configurations at C-8 and C-8', 1 was converted to an aryl-tetralin type compound (1a) with acetyl chloride by the previously reported reaction, in which inversion of the stereochemistry at C-7 of 3 to that of 3a was shown.<sup>19</sup> The observed spin coupling constants,  $J_{7,8} = 10.2$ Hz,  $J_{8,8'} = 10.5$  Hz and  $J_{8',7'ax} = 11.3$  Hz for **1a** confirmed the all-trans stereochemistry with two methyl groups and the pendant phenyl substituent all pseudo-equatorial positions (Figure 2).<sup>6</sup> Based on this evidence, 1 was determined to be (7S,8R,8'S)-3,4,3',4'-dimethylenedioxylignan-7-ol and named as saucerneol J.

Compound **2** was obtained as an colorless oil, with a molecular formula of  $C_{20}H_{20}O_6$  determined by HREIMS (*m/z*; found 356.1258 [M]<sup>+</sup>; calcd. 356.1260). The UV spectra of **2** revealed the presence of phenolic groups (244 and 291 nm). It showed the value of  $[a]_D^{22} + 21.9^\circ$  (*c* 0.08, CHCl<sub>3</sub>). The <sup>1</sup>H-NMR spectrum showed the presence of two methyl doublets (H-9 and H-9'), two methine groups (H-8' and H-8), two benzylic methine groups substituted by oxygen (H-7' and H-7), two dioxymethylene groups and five aromatic protons (H-2, H-5, H-6, H-3' and H-6'). The positions of each



Figure 1. Chemical structures of compounds 1-3

Notes



Figure 2. Partial structure of compound 1a.



Figure 3. Key HMBC correlations of compounds 1-2.

functional group were determined by a HMBC experiment. One set of correlations was observed between the oxygenated methine H-7 ( $\delta$  4.60) and C-2, C-6 and C-9 and the other set was between H-7' ( $\delta$  4.70) and C-2', C-6' and C-9'. Additionally, the correlations of phenolic hydroxyl signal at  $\delta$  8.19 were observed among the C-1', C-2' and C-3'. The relative stereochemistry of the tetrahydrofuran ring could be deduced by comparison with literature data on related lignans.<sup>2,20-26</sup> In the <sup>1</sup>H-NMR data, upfield-shifted signals of H-9 and H-9' at  $\delta$ 1.07 and 1.06, respectively, indicated *trans* positions to those methyl groups because if those two protons were located on cis position to those of methyl group, chemical shift values of two protons should be about 0.7 ppm due to the shielding effect of the aromatic rings.<sup>11,12</sup> Along with this, coupling constants of 9.2 and 10.0 Hz for H-7 and H-7', respectively, suggested trans-configurations both between H-7 and H-8 and between H-7' and H-8'.<sup>5,7</sup> From the above evidence, 2 was confirmed as  $(7\alpha, 8\beta, 7'\beta, 8'\alpha)$ -2'-hydroxy-3,4,4',5'-dimethylenedioxy-7,7'-epoxy lignan and named as saucerneol K.

## **Experimental Section**

General Experimental Procedures. Optical rotations were measured using a JASCO DIP-1000 (Tokyo, Japan) automatic digital polarimeter. The NMR spectra were recorded on a Bruker 250 MHz (DMX 250, Germany) and Varian 400 MHz (INOVA-400, USA) spectrometer. Samples were dissolved in CDCl<sub>3</sub>-*d* or CD<sub>3</sub>OD, and chemical shifts were reported in ppm downfield from TMS. HIEIMS and HIFABMS were obtained on a JEOL JMS700 spectrometer (JEOL, Japan). The stationary phases used for column chromatography (Silica gel 60, 70-230 and 230-400 mesh and Lichroprep RP-18 gel, 40-63 µm, Merck) and TLC plates (Silica-gel 60  $F_{254}$  and RP-18  $F_{254s}$ , 0.25 mm, Merck) were purchased from Merck KGaA (Darmstadt, Germany). Spots were detected under UV radiation and by spraying with 10% H<sub>2</sub>SO<sub>4</sub>, followed by heating. (*R*)-(-)- $\alpha$ -methoxy- $\alpha$ -(trifluoromethyl)-phenyacetyl chloride [(*R*)-MTPA-Cl] and (*S*)-(+)- $\alpha$ -methoxy- $\alpha$ -(trifluoromethyl)phenyacetyl chloride [(*S*)-MTPA-Cl] were purchased from Aldrich (St. Louis, MO, USA; purity 99.0%). All other chemicals and solvents were of analytical grade, and used without further purification.

**Plant Material.** The roots of *Saururus chinensis* was purchased in February 2003 from a folk medicine market, "Yak-ryong-si", in Daegu, Republic of Korea. These materials were confirmed taxonomically by Professor Ki-Hwan Bae, Chungnam National University, Daejeon, Republic of Korea. A voucher specimen (YNSC2004) has been deposited at the College of Pharmacy, Yeungnam University.

Extraction and Isolation. The dried roots of Saururus chinensis (9.7 kg) was extracted with 70% MeOH three times by refluxing for 24 hr, respectively and the MeOH solution was evaporated to dryness (1.0 kg). The MeOH extract was suspended in H<sub>2</sub>O (1.4 L), and the resulting H<sub>2</sub>O layer was successively partitioned with *n*-hexane, EtOAc and BuOH (each  $1.4 L \times 3$ ). The EtOAc extract (130 g) was loaded on a silica gel column (60 × 12 cm, Silica-gel 70-230 mesh) and eluted by a stepwise gradient of *n*-hexane-EtOAc (100:0  $\rightarrow$ 0:100) and then EtOAc-MeOH (100:0  $\rightarrow$  0:100). The eluates (500 mL in each flask) were combined into 39 fractions (SCE1-39) on the basis of silica gel TLC. Fraction SCE19 (970 mg) was chromatographed on a reverse-phase column (4  $\times$  50 cm, LiChroprep RP-18) and eluted by a stepwise gradient of MeOH-H<sub>2</sub>O (60:40  $\rightarrow$  100:0) and then Fraction SCE19-10 (160 mg) was chromatographed on a Sephadex LH-20 column ( $3 \times 90$  cm, Sephadex LH-20) eluted with MeOH (5.0 L) to give 1 (90 mg). Fractions 26 (1.0 g) was chromatographed on a reverse-phase column (4  $\times$  50 cm, LiChroprep RP-18) and eluted by a stepwise gradient of MeOH-H<sub>2</sub>O (40:60  $\rightarrow$  100:0) and then Fraction SCE26-13 (15 mg) was preparative TLC ( $10 \times 10$  cm, 0.25 mm coated silica gel,  $CHCl_3:MeOH = 95:5$ ) to give 2 (5 mg).

(7S,8R,8'S)-3,4,3',4'-Dimethylenedioxylignan-7-ol (1): Oil;  $[a]_{D}^{20}$  -65.7° (c 1.806, CHCl<sub>3</sub>); UV (MeOH)  $\lambda_{max}$  (log  $\varepsilon$ ) 234 (3.92), 287 (3.79) nm; <sup>1</sup>H-NMR (CD<sub>3</sub>OD, 250 MHz)  $\delta$  6.77 (1H, d, J = 1.3 Hz, H-2), 6.69 (1H, dd, J = 8.0, 1.5 Hz, H-6),6.69 (1H, d, J = 8.0 Hz, H-5), 6.69 (1H, d, J = 8.0 Hz, H-5'), 6.68 (1H, d, J = 1.5 Hz, H-2'), 6.60 (1H, dd, J = 7.9, 1.4 Hz, H-6'), 5.89 (2H, s, -OCH<sub>2</sub>O-), 5.88 (2H, s, -OCH<sub>2</sub>O-), 4.25 (1H, d, J = 9.6 Hz, H-7), 2.46 (2H, m, H-7'), 2.39 (1H, m, H-7'))H-8'), 1.75 (1H, m, H-8), 0.81 (3H, d, J = 6.2 Hz, H-9'), 0.54 (3H, d, J = 7 Hz, H-9); <sup>13</sup>C-NMR (MeOD, 62.9 MHz)  $\delta$  149.1 (C-3), 148.9 (C-3'), 148.2 (C-4), 147.0 (C-4'), 140.1 (C-1), 136.7 (C-1'), 122.9 (C-6'), 121.6 (C-6), 110.3 (C-2'), 108.8 (C-5), 108.6 (C-5'), 107.9 (C-2), 102.2 (-OCH<sub>2</sub>O-), 102.0 (-OCH<sub>2</sub>O-), 77.8 (C-7), 44.1 (C-8), 43.0 (C-7'), 35.2 (C-8'), 13.1 (C-9'), 10.5 (C-9); HRFABMS *m/z* 343.1538 ([M+H]<sup>+</sup>, calcd. for C<sub>20</sub>H<sub>23</sub>O<sub>5</sub>, 343.1545).

(7*R*,8*R*,8'*S*)-3,4,4',5'-Dimethylenedioxy-2',7-cyclolignan (1a):  $[a]_{D}^{22}$  +65.7° (*c* 0.036, CHCl<sub>3</sub>); <sup>1</sup>H-NMR (CDCl<sub>3</sub>, 250 MHz)  $\delta$  6.72 (1H, d, *J* = 7.9 Hz, H-5), 6.60 (1H, dd, *J* = 7.9, 1.7 Hz, H-6), 6.50 (1H, s, H-6'), 6.50 (1H, d, *J* = 1.6 Hz, H-2), 6.14 (1H, s, H-3'), 5.91 (2H, s, -OCH<sub>2</sub>O-), 5.80 and 5.79 (each 1H, d, J = 1.4 Hz, -OCH<sub>2</sub>O-), 3.36 (1H, d, J = 10.2 Hz, H-7), 2.69 (1H, dd, J = 16.1, 4.5 Hz, H-7'a), 2.54 (1H, dd, J = 16.3, 11.3 Hz, H-7'b), 1.60 (1H, m, H-8'), 1.44 (1H, ddq, J = 10.5, 10.5, 6.2 Hz, H-8), 1.03 (3H, d, J = 6.3 Hz, H-9'), 0.84 (3H, d, J = 6.2 Hz, H-9); <sup>13</sup>C-NMR (CDCl<sub>3</sub>, 62.9 MHz)  $\delta$  147.7 (C-3), 145.8 (C-4), 145.5 (C-5'), 145.4 (C-4'), 140.5 (C-1), 133.4 (C-2'), 130.1 (C-1'), 122.8 (C-6), 109.6 (C-3'), 109.1 (C-2), 107.7 (C-5), 107.6 (C-6'), 100.8 (-OCH<sub>2</sub>O-), 100.5 (-OCH<sub>2</sub>O-), 54.6 (C-7), 43.8 (C-8), 39.4 (C-7'), 35.4 (C-8'), 19.9 (C-9'), 17.1 (C-9).

(7*α*,8*β*,7'*β*,8'*α*)-2'-Hydroxy-3,4,4',5'-dimethylenedioxy-7,7'-epoxy Lignan (2): Colorless oil;  $[a]_{22}^{22}$  +21.9° (*c* 0.08, CHCl<sub>3</sub>); UV (CHCl<sub>3</sub>)  $\lambda_{max}$  (log  $\varepsilon$ ) 244 (3.24), 291 (3.01) nm; <sup>1</sup>H-NMR (CDCl<sub>3</sub>, 400 MHz)  $\delta$  8.19 (1H, s, OH), 6.87 (1H, d, J = 1.2 Hz, H-2), 6.81 (1H, dd, J = 6.5, 0.8 Hz, H-6), 6.79 (1H, d, J = 6.5 Hz, H-5), 6.45 (2H, s, H-3' and H-5'), 5.96 (2H, s, -OCH<sub>2</sub>O-), 5.89 (2H, s, -OCH<sub>2</sub>O-), 4.70 (1H, d, J = 10.0 Hz, H-7'), 4.60 (1H, d, J = 9.2 Hz, H-7), 1.98 (1H, m, H-8'), 1.76 (1H, m, H-8), 1.07 (3H, d, J = 6.4 Hz, H-9), 1.06 (3H, d, J =6.4 Hz, H-9'); <sup>13</sup>C-NMR (CDCl<sub>3</sub>, 100 MHz)  $\delta$  151.0 (C-2'), 148.2 (C-4), 147.9 (C-3), 147.5 (C-4'), 140.8 (C-1), 119.9 (C-6), 115.4 (C-1'), 108.4 (C-5), 106.9 (C-6'), 106.6 (C-2), 101.3 (-OCH<sub>2</sub>O-), 101.2 (-OCH<sub>2</sub>O-), 99.7 (C-3'), 89.5 (C-7'), 87.5 (C-7), 50.6 (C-8), 48.4 (C-8'), 14.1 (C-9' and C-9); HREIMS *m/z* 356.1258 ([M]<sup>+</sup>, calcd. for C<sub>20</sub>H<sub>20</sub>O<sub>6</sub>, 356.1260).

**Preparation of (S) and (R)-MTPA Esters of 1.** Mosher's esters was prepared according to the reported method.<sup>16-18</sup> To compound 1 (3 mg) in 0.5 mL of CH<sub>2</sub>Cl<sub>2</sub> were added sequentially 0.2 mL of anhydrous pyridine, 0.5 mg of 4-(dimethylamino)pyridine and 12.5 mg of (*R*)-(-)- $\alpha$ -methoxy- $\alpha$ -(trifluoromethyl)phenylacetyl chloride [(*R*)-MPTA-Cl]. The mixture was left at room temperature overnight and checked by TLC to determine if the reaction mixture was passed through a column (6 × 0.6 cm, silica gel, 230-400 mesh, 9385) with hexane-CH<sub>2</sub>Cl<sub>2</sub> (1:2). The eluate was dried in *vacuo* to give the (*S*)-MTPA ester of 1. Using (*S*)-MTPA-Cl, the (*R*)-MTPA ester of 1 was prepared.

**Conversion of 1 to 1a.** Compound **1** (10 mg) in 0.5 was dissolved in acetyl chloride (3 drops). The solution was kept at room temperature for 2 hr, and, after the addition of H<sub>2</sub>O, neutralized with aqueous NaHCO<sub>3</sub> and extracted with CH<sub>2</sub>Cl<sub>2</sub>. The organic layer was dried (Na<sub>2</sub>SO<sub>4</sub>), filtered and evaporated. The residue that dissolved in CH<sub>2</sub>Cl<sub>2</sub> (1-2 mL) was passed through a column ( $6 \times 0.6$  cm, silica gel, 230-400 mesh, 9385) with CH<sub>2</sub>Cl<sub>2</sub> mobile phase. The eluates were dried in *vacuo* to give compound **1a** (6 mg).

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