

Figure 1.

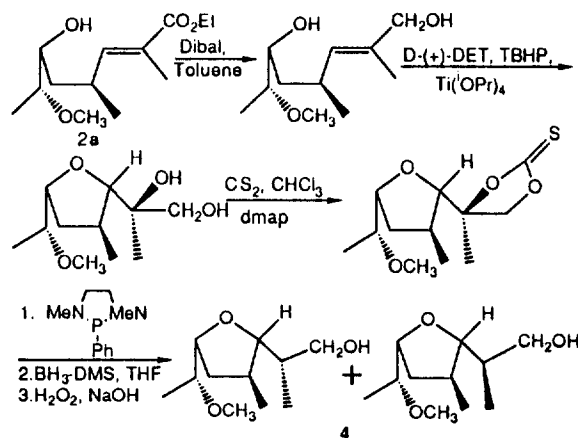
the stereochemistry at C-4 of α,β -unsaturated esters governs the stereochemical outcome for the formation of tetrahydrofuran esters. The rationale was also supported by the fact that even when there was no γ -substituent for α,β -unsaturated carboxylic ester such as the case for ring closure reaction toward nonactic acid, the predominant product was known to have 2,3-*cis* relationship⁷. Also similar stereochemical result can be found for the intramolecular conjugate addition of carbamate group in the α,β -unsaturated esters⁸. But it is interesting that when we prepared *Z* isomer of **2a** and carried out the cyclization to examine the effect of γ -substituent on stereoselectivity, we had the same stereochemical results as that of *E* isomer of **2a**. On the contrary to the previous report that the stereoselectivity for the conjugate addition of nucleophile to Michael acceptors depends upon *Z/E* configurations of Michael acceptors⁹, our results imply that the stereochemical induction for Michael acceptor was insensitive to *Z/E* configurations in our system. Perhaps $A^{1,3}$ strain due to the ester group was almost same as that due to 2-methyl group and thus the steric effect of γ -substituent became dominant in the process of stereochemical induction. However, it is still open to the question that the how much the steric effect of α -substituent of *Z/E* isomers can contribute to the stereochemical outcome along with that of competing γ -substituent in the intramolecular Michael reactions leading to tetrahydrofuran rings.

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- The structure of **3a** was determined by the chemical conversion of **2a** to tetrahydrofuran **4** through Sharpless asymmetric epoxidation-cyclization and the deoxygenation at

tertiary alcohol according to Corey's procedure (E. J. Corey and P. B. Hopkins, *Tetrahedron Lett.*, **23**, 1979 (1982).) and comparison of ¹H-NMR spectrum of **4** with those of the reduced forms of **3a**.



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Synthesis of Specifically Deuterated DNA Hexamer

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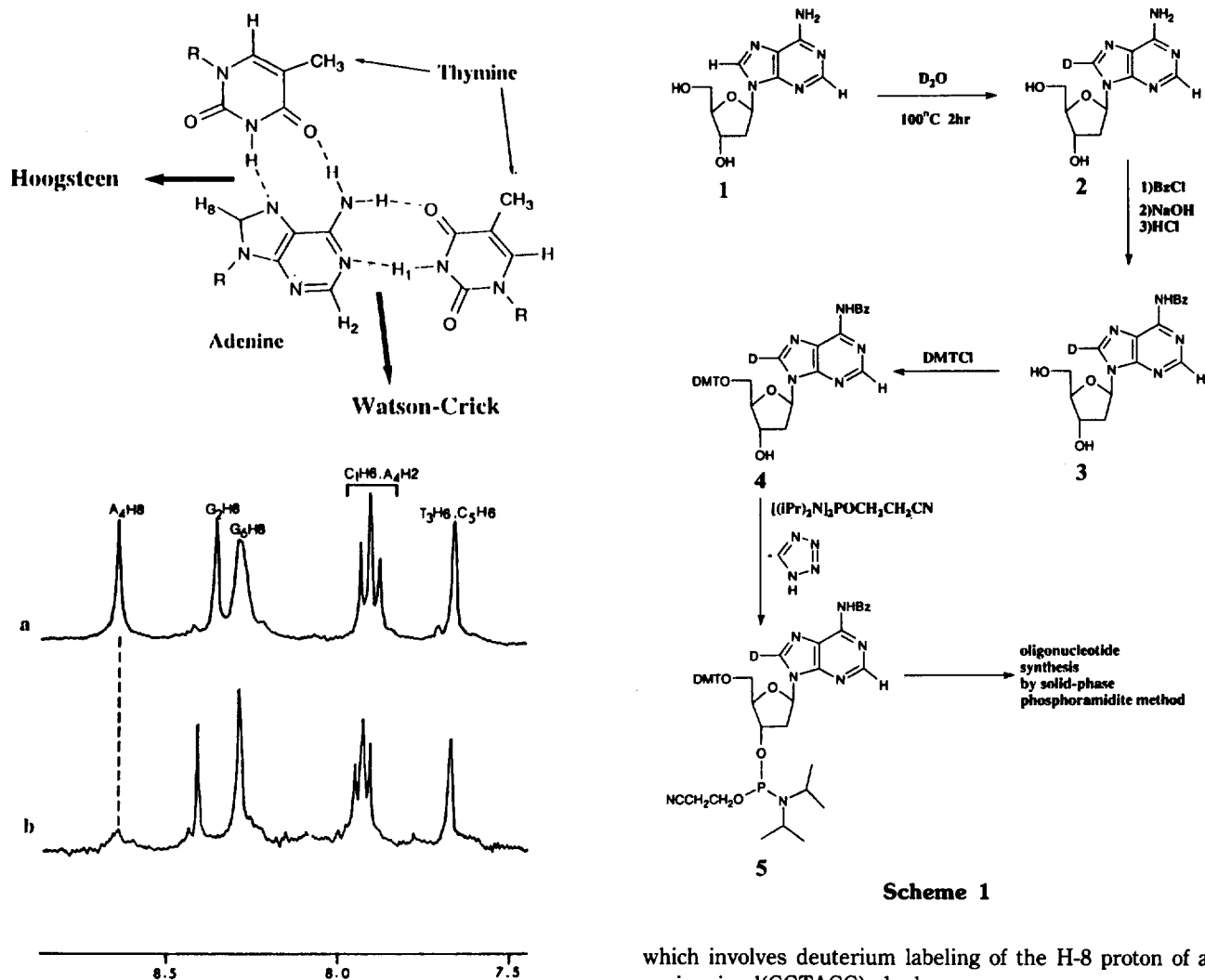
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The quinoxaline family of antibiotics of which echinomycin is a member are powerful antimicrobial and antitumor agents. The single-crystal X-ray study¹ of a echinomycin complex with d(CGACG) has shown that, surprisingly, the two central A-T base pairs are of the Hoogsteen type (Figure 1). Two-dimensional NMR studies of echinomycin complexes with d(ACGT) and d(ACGTACGT) duplexes have been reported^{2,3}. The van der Waals contacts detected in X-ray crystallographic analysis of are echinomycin-oligonucleotide co-

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Scheme 1

Figure 1. (a) 500 MHz $^1\text{H-NMR}$ spectrum of the aromatic proton resonances of the $d(\text{CGTACG})$ duplex obtained in D_2O buffer, pH 7.0, 25°C . Assignments of some of the DNA resonances are indicated on the figure. (b) Spectrum of DNA hexamer deuterated at the adenine H-8 position; otherwise similar to (a) Dotted line connect resonance position of the adenine H-8 proton. (Inset) Standard A. T and Hoogsteen A. T base pairs. The imino, H-2, and H-8 protons are designated H_1 , H_2 , and H_8 , respectively.

mplex are retained and the adenosine in the oligonucleotide adopts a syn orientation in solution, which is consistent with the formation of an A. T Hoogsteen base pair.

However, those previous NMR results have not provided definitive discrimination between Watson-Crick and Hoogsteen base pairs due to superposition of quinoxaline protons with the adenosine H-8 and H-2 protons. The present work would like to resolve this ambiguity. Isotopic labeling is the method of choice for NMR identification. Standard A. T base pairs have an adenine H-2 proton adjacent to the imino proton. In contrast, Hoogsteen base pairs have an adenine H-8 proton next to the imino proton (Figure 1). If a NOE from imino proton to carbon proton is observable, then that carbon can be labeled with deuterium in order to identify the NOE mate. Here we report a new synthetic methodology

which involves deuterium labeling of the H-8 proton of adenosine in $d(\text{CGTACG})$ duplex.

The strategies for labeling the adenine H-8 position are based on the fact that the proton at this position is labile at high temperature, both for adenine and for purines in nucleic acids⁴. Deoxyadenosine was deuterated as follows: 1 g of it was mixed with 100 g of D_2O and heated to 100°C for 2 hr in order to label the H-8 position. The mixture was cooled to 5°C overnight and the precipitate was removed by centrifugation. Deuteration was checked by NMR, and the optical absorption spectrum was unchanged. When the deuterium labeled deoxyadenosine was prepared, the protection of 6- NH_2 and 5'-OH and the phosphorylation of 3'-OH was carried out according to Scheme 1⁵.

The protection of the 6- NH_2 of adenine base and the 5'-OH of sugar moiety was achieved by the benzylation (step 2 \rightarrow 3) and dimethoxytritylation (step 3 \rightarrow 4), respectively. After phosphorylation, the adenine H-8 deuterated DNA hexamer $d(\text{CGTA}(\text{D})\text{CG})$ was synthesized *via* the solid phase phosphoramidite method⁶. NMR spectra of the aromatic region of the unlabeled and labeled DNA hexamer are shown in Figure 1. Assignments of some aromatic resonances are indicated on the spectra. We have found the adenine H-8 proton peak at 8.34 ppm which is definitely missing in the deuterated sample.

In conclusion, this method will provide a clue to clarifying above mentioned ambiguity. We are presently studying the

