# Sizes and Structures of Micelles of Cationic Octadecyl Trimethyl Ammonium Chloride and Anionic Ammonium Dodecyl Sulfate Surfactants in Aqueous Solutions

Hong-Un Kim<sup>†</sup> and Kyung-Hee Lim<sup>\*</sup>

Department of Chemical Engineering, Chung-Ang University, Seoul 156-756, Korea Received May 9, 2003

The sizes and structures of micelles formed in aqueous solutions of cationic octadecyl trimethyl ammonium chloride (OTAC) and anionic ammonium dodecyl sulfate (ADS) surfactants were investigated using small-angle neutron scattering (SANS), self-diffusion coefficients by pulsed-gradient spin-echo (PGSE) NMR, and dynamic light scattering (DLS) methods. SANS and DLS data indicate that their structures are spherical at concentrations as high as 300 mM. As the total surfactant concentration increases, the peaks of SANS spectra shift to higher scattering vector and become sharper, indicating that the intermicellar distance decreases and its distribution becomes narrower. This is due to more compact packing of surfactant molecules at high concentrations. The intermicellar distance of around 100 Å above 200 mM corresponds approximately to the diameter of one micelle. The sizes of spherical micelles are 61 Å and 41 Å for 9 mM OTAC and 10 mM ADS, respectively. Also the self-diffusion coefficients by PGSE-NMR yield the apparent sizes 96 Å and 31 Å for micelles of 1 mM OTAC and 10 mM ADS, respectively. For ADS solutions of high concentrations (100-300 mM), DLS data show that the micelle size remains constant at  $25 \pm 2$  Å. This indicates that the transition in micellar shape does not take place up to 300 mM, which is consistent with the SANS results.

**Key Words :** Ammonium dodecyl sulfate (ADS), Octadecyl trimethyl ammonium chloride (OTAC), Smallangle neutron scattering (SANS), Pulsed-gradient spin-echo (PGSE)-NMR, Intermicellar distance

#### Introduction

One of the characteristics of surfactants is the capacity to form association structures in solutions. Micelles are one type of such structures, and the minimum concentration to form micelles is called the critical micelle concentration (CMC). The CMC is not a specific concentration but a narrow range of concentrations. The micellar solutions above CMC are homogeneous and transparent, thus it is difficult to observe size and structure of micelles. In general, it is known that the size of micelles formed in aqueous surfactant solution ranges from 10 to 100 Å, and that the shape of micelles varies with increasing concentration in order of spherical, cylindrical, hexagonal, and lamellar in aqueous ionic surfactant solutions.<sup>1-4</sup>

Small-angle neutron scattering (SANS) can probe interfacial structures of surfactant aggregations, polymers, and biomolecules, utilizing the neutron scattering intensity difference between hydrogen and deuterium. A good permeability and no charge of neutrons favor internal structure analysis. Therefore, SANS has been widely used for the determination of sizes and structures of micelles.

Pulsed-gradient spin-echo (PGSE) NMR spectrometry has the ability to measure the self-diffusion coefficients of monomers and micelles in aqueous surfactant solutions, which thus enables to probe the interactions between them.<sup>5</sup> The formation of micelles and its phase transition in aqueous surfactant solutions is related to the diffusion of hydrophobic parts in the micellar phase. Thus, the self-diffusion coefficient is a measure of analyzing the molecular organization in solutions and is highly affected by the changes in structure and association/dissociation in colloidal dispersions.<sup>6</sup> The self-diffusion coefficient ( $D_s$ ) in aqueous surfactant solutions decreases with increasing surfactant concentration, which is due to the hindrance of diffusion by interaction between surfactant monomers. Also self-diffusion of micelles proceeds slowly since the size of micelles is larger than that of monomers. As the surfactant concentration increases, the transition of shape from spherical to cylindrical contributes to the reduction of self-diffusion coefficient, too.

We reported previously the CMC, its dependence on temperature, and thermodynamics of micellization for the cationic octadecyl trimethyl ammonium chloride (OTAC) and anionic ammonium dodecyl sulfate (ADS).<sup>7</sup> OTAC has been widely used in industries due to excellent antistatic effects and softness, and ADS has been used in cosmetics and personal cares in place of the well-known sodium dodecyl sulfate (SDS) which undergoes autocatalytic acid hydrolysis in acidic solutions or at high temperatures and causes skin irritation. No information on sizes and structures of micelles at different concentrations has been obtained for these surfactant systems.

The sizes, shapes, and transition of OTAC and ADS micelles in aqueous solutions have been investigated using SANS, self-diffusion coefficient by PGSE-NMR, and dynamic light scattering (DLS), and their results are reported in this article.

<sup>\*</sup>Corresponding Author. Tel: +82-2-820-5275, Fax: +82-2-826-3574; e-mail: khlim@cau.ac.kr

<sup>&</sup>lt;sup>†</sup>Present address: Department of Applied Chemistry, Dongyang Technical College, Seoul 152-714, Korea

Sizes and Structures of Micelles of OTAC and ADS Surfactants

### **Experimental Section**

**Materials.** The anionic ammonium dodecyl sulfate (ADS) and cationic octadecyl trimethyl ammonium chloride (OTAC) surfactants were purchased from Fluka. ADS was in the form of a 30% aqueous solution. It was placed in a rotary evaporator to reduce the contents of water and volatile materials. Then it was recrystallized with 90% ethanol and absolute ethanol twice, respectively. The OTAC had a stated purity of 98% and was recrystallized three times with absolute ethanol. The purified surfactants were finally dried in an evacuated desiccator.<sup>8</sup> All samples for SANS and PGSE-NMR measurements were prepared in D<sub>2</sub>O (Aldrich, purity 99.9 atom % D).

**Small-angle neutron scattering (SANS).** The fundamental principles of SANS are that the scattering vector, q, corresponds to the change in neutron momentum and that the scattered intensity,  $I_s(q)$ , changes with the sample. The scattering vector is defined as

$$q = \left| \vec{q} \right| = \frac{4\pi}{\lambda} \sin\!\left(\frac{\theta}{2}\right) \tag{1}$$

where  $\lambda$  and  $\theta$  are the wavelength and scattered angle of neutron. The intensity of neutron scattered is given by

$$I_{s}(q) = \Phi A dT_{S+SB} \left( \frac{d\Sigma}{d\Omega} (\vec{q}) \right)_{S} \Delta \Omega \varepsilon t .$$
<sup>(2)</sup>

Here,  $\Phi$  is the neutron flux unto the sample, *A* sample area, *d* sample thickness,  $T_{S+SB}$  transmission through the sample plus sample cell,  $\Sigma$  the scattering cross area,  $\Omega$  the solid angle subtended by one pixel of the detector,  $\varepsilon$  detector efficiency, and *t* effective counting time, respectively. The absolute scattering cross area  $[d\Sigma(\vec{q})/d\Omega]_S$  includes the form factor  $P(\vec{q})$  and structure factor  $S(\vec{q})$  as well as number density  $(N_P)$  of particle as follows.

$$\left(\frac{d\Sigma}{d\Omega}(\vec{q})\right)_{S} = N_{P}P(\vec{q})S(\vec{q})$$
(3)

The scattering vector q and the neutron wavelength ranged from 0.006 to 0.6 Å<sup>-1</sup> and 4 to 8 Å, respectively. The resolution of wavelengths was 10% with full width at halfmaximum value. The samples prepared in  $D_2O$  were injected carefully without making bubbles into quartz cylindrical cells (Next Instrument Co.) with a path length of 2 mm and outside diameter of 22 mm. The calibration of absolute scattering intensity was carried out using silica and the scattering intensity was normalized by dividing by that of  $D_2O$ . The measured data were treated according to the procedures of Hanaro of Korea Atomic Energy Research Institute (KAERI).<sup>9</sup>

The scattering intensity  $I_s(q)$  measured of the sample and the Guinier law, equation (4), yield the radius of gyration  $(R_g)$ .<sup>10</sup>

$$I_s(q) = I(0) \exp\left(\frac{-R_g^2}{3}q^2\right) \quad \text{for } 0.5 < qR_g < 1.4$$
(4)

*Bull. Korean Chem. Soc.* **2004**, Vol. 25, No. 3 383

where  $I_s(q)$  and I(0) are the intensities of scattered neutrons with and without sample, respectively, which are functions of scattering vector q. The radius ( $R_{mic}$ ) of spherical micelles is calculated from the relation:<sup>11</sup>

$$R_{mic} = \sqrt{5/3}R_g. \tag{5}$$

The shape of micelles can be determined using *n* values of the power-law,  $I_s(q) \propto q^{-n}$ . The spherical micelle is identified when n = 4 according to Porod's law.<sup>12</sup> Also n = 1 points to the cylindrical or rod-like micelles at  $1/l_P < q < 1/R_{mic}$  where  $l_P$  is the length of cylindrical micelles.<sup>13,14</sup>

The length  $l_P$  can be determined from the Holtzer plot or bending rod plot in which  $qI_s(q)$  is plotted versus q.<sup>13,15,16</sup> That is,  $l_P$  is obtained from  $ql_P \approx 1.9$  for which the q value is determined from the intersection point of linear fits for low and high q values.

Measurements of self-diffusion coefficients by PGSE-NMR. The self-diffusion coefficients ( $D_s$ ) of monomer and micelle were measured using the pulsed-gradient spin-echo (PGSE) NMR with a Bruker DMX 600 spectrometer. The echo attenuation intensity A(g) as a function of gradient amplitude (g) is expressed by equation (6), as was found to decay exponentially.

$$A(g) = A(0)\exp[-\gamma^2 \delta^2 (\Delta - \delta/3) D_s g^2]$$
(6)

Here, A(0) is the echo intensity in the absence of gradient,  $\gamma$  is the gyromagnetic ratio of proton, whose value is given by  $2.675 \times 10^8 \text{ T}^{-1} \text{s}^{-1}$ .<sup>17</sup> Also  $\delta$  and  $\Delta$  are pulsed gradient duration and the time interval between two gradient pulses, respectively, as depicted in Figure 1.<sup>18-20</sup> In Figure 1, it is shown that two radio frequency pulses and two field gradient pulses of magnitude *g*, duration  $\delta$ , separation  $\Delta$  were applied before and after the 180° refocusing pulse. The second half of the spin echo was Fourier-transformed, and the relative intensities of the resolved absorption peaks were measured.

Values of  $D_s$  were determined from the slope of  $\ln(A(g)/A(0))$  versus  $g^2$  plots at fixed  $\delta$  and  $\Delta$  with varying g in equation (6).<sup>21,22</sup> Calibration was carried out with  $D_2O$  and its self-diffusivity was measured to be  $1.865 \times 10^{-9} \text{ m}^2/\text{s}$ , which is in excellent agreement with the literature value,  $1.872 \times 10^{-9} \text{ m}^2/\text{s}$ .<sup>23</sup> Based on this calibration result,  $D_s$  of methanol at 25 °C was determined as  $1.98 \times 10^{-9} \text{ m}^2/\text{s}$  from the slope -31.164 for  $\delta = 4$  ms and  $\Delta = 15.1$  ms. This value is

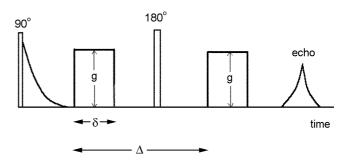
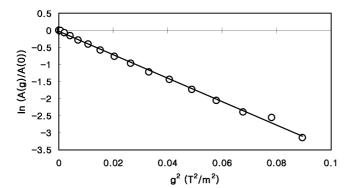


Figure 1. Schematic of the pulsed-gradient spin-echo pulse sequence.



**Figure 2**. Plot of  $\ln(A(g)/A(0))$  versus  $g^2$  for the determination of  $D_s$  at 25 °C for 1 mM ADS solution. The diffusivity  $D_s$  is calculated from the slope (= -34.367); the correlation coefficient is 0.9987.

compared well with that of literature,  $2.30\times 10^{-9}\,m^2\!/s.$   $^{24,25}$ 

Figure 2 shows an example of how to determine  $D_s$  from equation (6) for the 1 mM ADS solution. The self-diffusion coefficients of monomers,  $D_s^{mon}$ , and of micelles,  $D_s^M$ , were measured directly at concentrations below and above the CMC by PGSE-NMR. The NMR peaks characteristic to the surfactant was observed conspicuously at these concentrations.

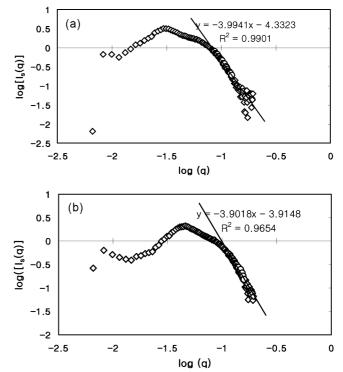
**Dynamic light scattering (DLS).** In order to determine the sizes and their distribution of OTAC and ADS micelles, scattered dynamic light intensities were measured using a ZetaSizer 1000HS (Malvern Instruments Ltd, UK) with a capillary cell. Brownian motions of the micelles were observed by DLS and related to their sizes.

The detector angle of 90 degrees, temperature of 25 °C, and wavelength of 633 nm were set. The refractive indices of OTAC (1.44), ADS (1.37), and solvent deionized (D.I.) water (1.33), and the viscosity of solvent (0.891 cP) were used. All samples were prepared in D.I. and triply distilled water.

#### **Results and Discussion**

Size and structure of micelles by SANS. The SANS spectra of 9 mM OTAC and 10 mM ADS solutions in  $D_2O$  were measured and the plots of  $\log(q)-\log(I_s(q))$  are made. One of such plots is shown in Figure 3. The slopes at high q's are -3.994 and -3.902, respectively. These values are very close to -4, which signifies that the Porod's law,  $I_s(q) \propto q^{-4}$  is satisfied. These results point to the fact that the micelles in the aqueous 9 mM OTAC and 10 mM ADS solutions are spherical.

Surfactant association structures may be explained by a geometric analysis, *i.e.*, by the values of critical packing parameter,  $N_{P_c} = V/a_0 l$ .<sup>26</sup> Here V and l are the volume and length of the alkyl chain of the surfactant and they are usually calculated from  $V = (27.4+26.9N_c)$  Å<sup>3</sup> and  $l = (1.54+1.265N_c)$  Å × 0.77 with  $N_c$  being the carbon number in the hydrophobic chain of the surfactant.<sup>27</sup> The factor 0.77 in the length l reflects that the real length is essentially 77% of fully extended length of the carbon chain. The quantity  $a_0$ 



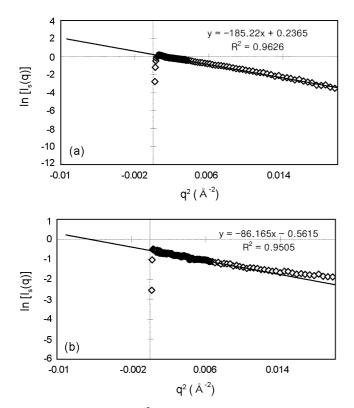
**Figure 3**. Plot of log(q) versus  $log(I_s(q))$  and their linear fits at high q for 9 mM OTAC (a) and 10 mM ADS (b).

is the area of the surfactant head group and is determined from the slope of surface tension versus concentration,  $\gamma$ – log*C*, plots. For OTAC and ADS,  $N_p$  values are 0.2 and 0.37 calculated from the  $\gamma$ –log*C* data.<sup>28</sup> These values suggest that the OTAC and ADS molecules be conical in shape and that, since  $N_{P_c} < 1/3$  or  $N_p \sim 1/3$ , their micelles be spherical in aqueous solutions,<sup>29</sup> which is in good agreement with SANS data.

In order to determine the sizes of micelles, plots of  $q^2$  versus  $\ln(I_s(q))$  were made and one example of such plots is shown in Figure 4. The radii of gyration for 9 mM OTAC and 10 mM ADS are obtained as 23.6 Å ( $0.6 < qR_g < 1.4$ ) and 16 Å ( $0.5 < qR_g < 1.3$ ),<sup>10</sup> respectively, from the slopes of linear fits (Figure 4). The radii of spherical micelles ( $R_{mic}$ ) are then determined as 30.5 and 20.7 Å for 9 mM OTAC and 10 mM ADS solutions, respectively, from  $R_{mic} = \sqrt{5/3} R_g$ . The radius (20.7 Å) of ADS spherical micelles is larger than that (18 Å) of the sodium dodecyl sulfate (SDS) measured by SANS at dilute aqueous solutions.<sup>30,31</sup> In ADS and SDS micelles, the counterion (NH<sub>4</sub><sup>+</sup>) in the former is larger than that (Na<sup>+</sup>) in the latter. This is why the size of the ADS micelles is larger.

The sizes and structural transitions of ADS and OTAC micelles were examined by SANS at various surfactant concentrations and the SANS results are shown in Figure 5. Both surfactant systems exhibit peaks in intensity, which means, according to Gräbner *et al.*,<sup>32</sup> that spherical micelles were formed at the concentrations examined. If the shape of the micelles were cylindrical, plateaus (instead of peaks) would appear in the SANS spectra.<sup>32,33</sup> Hence the results of

Sizes and Structures of Micelles of OTAC and ADS Surfactants

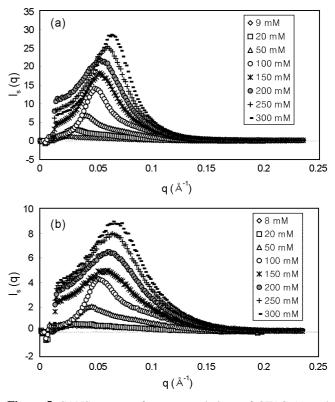


**Figure 4**. Guinier plot of  $q^2$ -ln( $I_s(q)$ ) and determination of gyration radius for 9 mM OTAC (a) and 10 mM ADS (b).

Figure 5 imply that no structural transition from spherical to cylindrical micelles was observed up to 300 mM. In contrast, such transition was observed at 250 mM for SDS micelles, examined by X-ray scattering.<sup>34</sup>

The locations of the peaks correspond to the intermicellar distance. The position of peaks shifts toward high q, as surfactant concentration increases. This result indicates that the intermicellar distance (calculated from  $2\pi/q_{peak}$ ) becomes smaller, as repulsive interactions between micelles become stronger at higher concentrations. On the other hand, the micelle sizes change little with surfactant concentrations up to 50 mM. The micelle sizes and intermicellar distances at various surfactant concentrations are presented in Table 1.

With increasing surfactant concentrations, the intermicellar



**Figure 5**. SANS spectra of aqueous solutions of OTAC (a) and ADS (b) at various surfactant concentrations.

distance decreases significantly up to 50 mM and then the rate of decrease is reduced substantially. Considering the diameters of a micelle (61 and 41 Å for OTAC and ADS micelles, respectively), the intermicellar distance at high surfactant concentrations correspond to the sizes of one or two micelles. Structural transition from spherical to cylindrical micelles may occur, when the head groups of neighboring micelles interact repulsively each other (Figure 6). However, for OTAC and ADS micelles, the intermicellar distances are large enough for micellar interactions to be weak and therefore for the structural transition not to occur.

The peaks become sharper with increasing surfactant concentration. This means that the distribution of intermicellar distance is narrower due to stronger repulsive

Table 1. Micelle sizes and intermicellar distances at various concentrations of cationic OTAC and anionic ADS surfactants

	OTAC			ADS		
Concentration (mM)	Diameter (Å)	Intermicallar distance (Å)	Concentration (mM)	Diameter (Å)	Intermicellar distance (Å)	
9	$61 \pm 4$	246	10	$41 \pm 4$	347	
20	$60 \pm 4$	213	20	$44 \pm 4$	201	
50	$60 \pm 6$	153	50	$44 \pm 4$	136	
100	_	127	100	_	119	
150	_	118	150	_	112	
200	_	121	200	_	101	
250	_	104	250	_	97	
300	_	99	300	_	97	

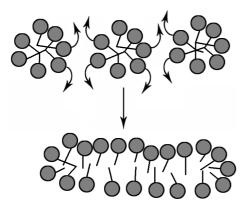


Figure 6. Schematic of structural transition from spherical to cylindrical micelles.

interaction among micelles when the solution becomes more crowded with micelles.<sup>33</sup>

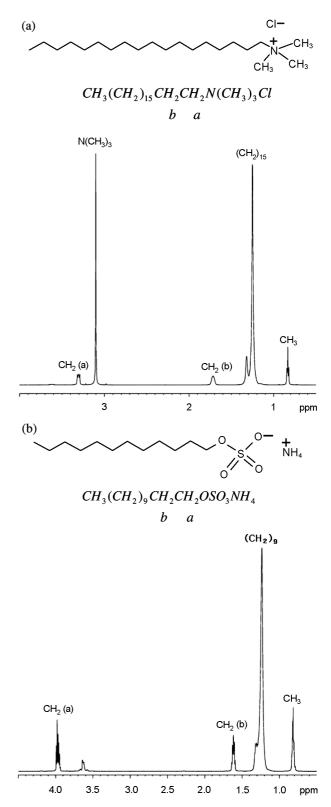
NMR spectra and determination of self-diffusion coefficients. Figure 7 shows the NMR spectra of 1 mM OTAC (CMC = 0.33 mM) and 10 mM ADS (CMC = 6.5mM) solutions above CMCs along with their respective structural formula. In Figure 7a the NMR peaks of OTAC were observed at 0.836 (CH<sub>3</sub> functional group), 1.213 ((CH<sub>2</sub>)<sub>15</sub> alkyl chain group), 1.69 (CH<sub>2</sub>(b)), 3.006 (N(CH<sub>3</sub>)<sub>3</sub>), 3.23 ppm (CH<sub>2</sub>(a)), respectively. Similarly for ADS, the peaks were observed at 0.836 (CH<sub>3</sub> functional group), 1.213 ((CH<sub>2</sub>)<sub>15</sub> alkyl chain group), 1.69 (CH<sub>2</sub>(b)), 3.995 ppm  $((CH_2(a)))$ , respectively, and a small peak at 3.62 ppm which corresponds to some impurities. At 0.2 mM OTAC and 1 mM ADS below CMC, NMR peak exhibits the same points as those of micelles. Therefore, we measured the selfdiffusion coefficient directly at the largest of these peaks except for that of alkyl chain group.

Based on the NMR data, the self-diffusion coefficients  $D_s$ 's of OTAC and ADS were determined at 3.006 ppm (N(CH<sub>3</sub>)<sub>3</sub>) and 0.836 ppm (CH<sub>3</sub>).  $D_s$ 's were calculated from the slopes of the plots of  $\ln(A(g)/A(0))$  versus  $g^2$ . Figure 8 shows such plots for OTAC (1 mM) and ADS (10 mM) micelles. The self-diffusivity  $D_s$ 's calculated from the slopes of the plots are presented in Table 2.

For ADS, the monomer self-diffusivity  $(D_s^{mon})$  was three times larger than the micelle diffusivity  $(D_s^M)$ , which reflects the fact that micelles are larger than monomers. For OTAC micelles the diffusivity  $D_s^M$  is order-of-magnitude smaller than that of ADS micelles. This result indicates that the OTAC micelles are larger than the ADS micelles.

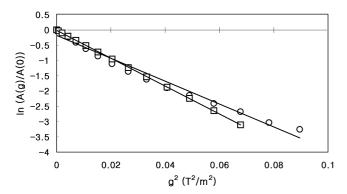
Notably, the OTAC monomer diffusivity measured at 0.2 mM is approximately the same as the OTAC micelle diffusivity measured at 1 mM. This indicates that  $D_s^{mon}$  at 0.2 mM reflects the diffusivities of many monomers and a few micelles. Previously we reported that CMC of OTAC at 25 °C was 0.324 mM from the conductivity data.<sup>7</sup> However, CMC is a narrow concentration range and depends on measurement method. The conductivity method may yield higher CMC, because micelles appeared to be formed at 0.2 mM OTAC according to the NMR data. To avoid such

Hong-Un Kim and Kyung-Hee Lim



**Figure 7**. Structural formula and NMR spectra for the aqueous solutions of (a) 1 mM OTAC and (b) 10 mM ADS.

discrepancy, NMR spectra should be obtained at lower surfactant concentrations. That is, when CMC is low, *i.e.*, below 1 mM,  $D_s^{mon}$  is usually determined from  $D_s^{obs}$  and equation (7)



**Figure 8**. Plot of  $\ln(A(g)/A(0))$  versus  $g^2$  for 1 mM OTAC (  $\Box$  ) and 10 mM ADS aqueous solutions (  $\bigcirc$  ) at 25 °C.

**Table 2.** Self-diffusion coefficients of monomers  $(D_s^{mon})$  and micelles  $(D_s^M)$  of OTAC and ADS in  $D_2O$  solutions

Self-diffusion coefficients	OTAC	ADS
$D_s^{mon}$ (× 10 <sup>-11</sup> m <sup>2</sup> /s)	6.63	42.37
$D_s^M (\times 10^{-11} \mathrm{m^2/s})$	4.24	15.08

$$D_{s}^{obs} = D_{s}^{M} + \frac{CMC}{C_{t}} (D_{s}^{mon} - D_{s}^{M})$$
(7)

where  $C_t$  is the total surfactant concentration.<sup>5</sup> However, for OTAC this method could not be applied because the NMR signal was too weak for dilute solutions at the prescribed chemical shifts.

As shown in Table 2, the self-diffusion coefficients of micelles are smaller than those of monomers as expected since the size of micelles is larger than that of monomers for both ionic surfactant solutions. The sizes of ADS monomer and micelles are smaller than those of OTAC, thus the self-diffusion coefficients for ADS monomer and micelles are larger than those for OTAC. Therefore, the values of self-diffusion coefficients for micelles provide the information on micelle sizes.

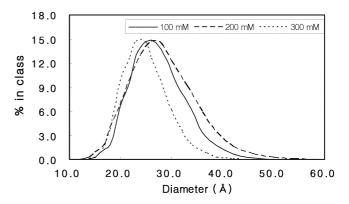
The apparent radius of micelles  $(R_{app})$  can be estimated from the Stoke-Einstein relation which may be expressed as

$$D_s = \frac{kT}{6\pi\eta R_{app}} \tag{8}$$

where k is the Boltzmann constant, T is the measurement temperature (298 K), and  $\eta$  is the viscosity of the medium, which is considered as almost the same as that of water (1.0 cP).

 $R_{app}$  of the OTAC micelle at 1 mM solution is 48 Å, which is a little larger than that by SANS (30.5 Å for 9 mM OTAC solution).  $R_{app}$  of the ADS micelles is 16 Å at 10 mM and is rather smaller than that by SANS (21 Å) at the same concentration. Hence, the self-diffusivity by PGSE-NMR yields sizes different from those by SANS and their difference may be significant (20-40%).

Micelle sizes by DLS. Figure 9 shows the size distribution of ADS micelles in aqueous solutions of high surfactant



**Figure 9**. Size distributions of ADS micelles determined by DLS at high surfactant concentrations.

concentrations, 100, 200, and 300 mM. Micelles formed at these high concentrations were monodispersed since their polydispersity ranged from 0.03 to 0.05. The average diameter of ADS micelles was estimated as  $25 \pm 2$  Å at these concentrations, which is approximately twice the 77% (12 Å) of the alkyl chain length of ADS. DLS yielded the smaller micelle size than SANS and PGSE-NMR. At high surfactant concentrations, smaller micelles were obtained probably because of increased interactions between micelles. DLS data also show that the shape transition of ADS micelles did not occur at concentrations up to 300 mM, which is in agreement with the SANS result.

Micelle sizes are determined by SANS for dilute solutions of the surfactant concentrations slightly above CMC to minimize interactions between micelles. However, DLS data are obtained often for concentrated surfactant solutions. For dilute solutions, DLS signals are often too weak to be analyzed for size determination.

## Conclusions

The sizes of micelles of cationic surfactant OTAC and anionic surfactant ADS in aqueous solutions have been measured by SANS, PGSE-NMR (self-diffusion coefficients), and DLS. Three methods yielded slightly different micelle diameters: for OTAC micelles, 61 Å (SANS), 96 Å (PGSE-NMR); for ADS micelles, 41 Å (SANS), 31 Å (PGSE-NMR), and 25 Å (DLS).

The shape of micelles is spherical, which is confirmed by the SANS data satisfying the Porod's law,  $I_s(q) \propto q^{-4}$ . The intermicellar distance with increasing surfactant concentration becomes smaller due to stronger repulsive interactions between micelles at high concentrations. Although the intermicellar distance becomes almost constant above 200 mM, the structural transition of spherical micelles to cylindrical ones was not observed up to 300 mM surfactant concentrations. DLS results also support for no structural transition of micelles over the concentrations examined.

Acknowledgements. This study was supported by Korea Institute of Science & Technology Evaluation and Planning 388 Bull. Korean Chem. Soc. 2004, Vol. 25, No. 3

(KISTEP) and Ministry of Science & Technology (MOST), Korean government through its National Nuclear Technology Program.

H. U. Kim was supported by Korea Research Foundation (Program No: EN0037) for this work. Advices by Dr. B.-S. Seong and Y.-S. Han of Hanaro, Korea Atomic Energy Research Institute for SANS, Dr. C.-J. Cheong and E.-H. Kim of Korea Basic Science Institute for PGSE-NMR, and Bomoon Trading Co. for DLS are greatly appreciated.

#### References

- 1. Courchene, W. L. J. Phys. Chem. 1964, 68, 1870.
- 2. Iwadare, Y.; Suzawa, T. Nippon Kagaku Zasshi 1969, 90, 1106.
- Benedough, D.; Chen, S.-H.; Koeler, W. C. J. Phys. Chem. 1983, 87, 2621.
- 4. Chevalier, Y.; Chachaty, C. Colloid Polym. Sci. 1984, 262, 489.
- Misselyn-Bauduin, A.-M.; Thibaut, A.; Grandjean, J.; Broze, G.; Jerome, R. J. Colloid Interface Sci. 2001, 238, 1.
- Walther, K. L.; Gradzielski, M.; Hoffmann, H.; Wakaun, A. J. Colloid Interface Sci. 1992, 153, 272.
- Kang, K.-H.; Kim, H.-U.; Lim, K.-H. Colloid Surf. A 2001, 189, 113.
- Armarego, W. L. F.; Perrin, D. D. In *Purification of Laboratory Chemicals*, 4th ed.; Butterworth-Heinemann: Oxford, 1996.
- 9. Korea Atomic Energy Research Institute, In *Computing Guide for* Hanaro Small-Angle Scattering Data Treatment, 2002.
- Guinier, A.; Fournet, G. In Small-Angle Scattering of X-Rays; John Wiley & Sons: New York, 1955.
- Feigin, L. A.; Svergun, D. I. In *Structure Analysis by Small-Angle X-Ray and Neutron Scattering*; Taylor, G. W., Ed.; Plenum Press: New York and London, 1987; pp 68-69.
- Porod, G. In Small-Angle X-Ray Scattering; Glatter, O.; Kratky, O., Ed.; Academic Press: New York, 1982; p 17.

- Hong-Un Kim and Kyung-Hee Lim
- Pedersen, J. S.; Egelhaaf, S. U.; Schurtenberger, P. J. Phys. Chem. 1995, 99, 1299.
- Lin, M. Y.; Hanley, H. J. M.; Sinha, S. K.; Straty, G. C.; Peiffer, D. G.; Kim, M. W. *Physica B* 1995, *213-214*, 613.
- 15. Holtzer, A. J. Polym. Sci. 1995, 17, 432.
- 16. Denkinger, P.; Burchard, W. J. Polym. Sci. B, Polym. Phys. 1991, 29, 589.
- 17. Kato, T. J. Phys. Chem. 1985, 89, 5750.
- Braun, S.; Kalinowski, H.-O.; Berger, S. In 100 and More Basic NMR Experiments; A Practical Course; VCH: 1999; Ch.11, pp 349-351.
- Lindman, B.; Olsson, U.; Soderman, O. In *Handbook of Micro-emulsion Science and Technology*; 1998; Ch.10, pp 309-356.
- 20. Walderhaug, H.; Nystrom, B. Colloid Surf. A 1999, 149, 379.
- Annunziata, O.; Costantino, L.; D'Errico, G.; Paduano, L.; Vitagliano, V. J. Colloid Interface Sci. 1999, 216, 16.
- Caria, A.; Regev, O.; Khan, A. J. Colloid Interface Sci. 1998, 200, 19.
- 23. Bruker Co., Almanac, 2001; p 72.
- 24. Weingartner, H. Z. Phys. Chem. (Neue Folge) 1982, 132, 129.
- 25. Holz, M.; Weingartner, H. J. Magn. Reson. 1991, 92, 115.
- Israelachvili, J. N. In *Intermolecular and Surface Forces*, 2<sup>nd</sup> Ed.; Academic Press: London, 1991.
- 27. Tanford, C. J. Phys. Chem. 1974, 78, 2469.
- 28. Kim, H.-U. Ph.D Dissertation; Chung-Ang University: Korea, 2002.
- Jonsson, B.; Lindmann, B.; Holmberg, K.; Kronberg, B. In Surfactants and Polymers in Aqueous Solutions; John Wiley and Sons: New York, 1997; pp 84-100.
- 30. Cabane, B.; Duplessix, R.; Zemb, T. J. Phys. 1985, 46, 2161.
- Førland, G. M.; Sameth, J.; Gjerde, M. I.; Hfiland, H.; Jensen, A. F.; Mortensen, K. J. Colloid Interface Sci. 1998, 203, 328.
- 32. Grabner, D.; Matsuo, T.; Hoinkis, E.; Thunig, C.; Hoffmann, H. J. Colloid Interface Sci. 2001, 236, 1.
- Kim, H. U. Ph. D. Dissertation; Chung-Ang University: Seoul, Korea, 2002; p166.
- 34. Reiss-Husson, F.; Luzzati, V. J. Phys. Chem. 1964, 86, 3504.