## Bioisoster of Capsaicin: Synthesis of 1-Hydroxy-2-pyridone Analogue

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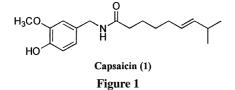
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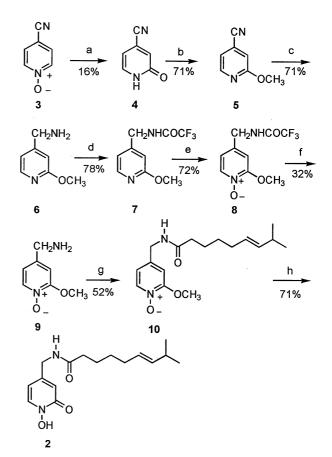
After capsaicin (1), a pungent principle of *capsicums*,<sup>1</sup> was known to act on a subset of peripheral sensory neuron,<sup>2</sup> molecular approach toward more potent capsaicin agonists which are anticipated to be useful as novel analgesic agents has been extensively studied.<sup>3</sup> Three parts of capsaicin structure - the aromatic ring region, the amide bond region, and the side chain region - were modified and it was found that the catechol moiety in capsaicin is important for the analgesic activity.<sup>3a</sup>

By application of bioisosterism, we previously found that the analogue which replaced the catechol moiety in dopamine with 1-hydroxy-2-pyridone system also revealed the similar dopaminergic activity.<sup>4</sup> Based on our result, we now replaced the functional catechol moiety in capsaicin with 1hydroxy-2-pyridone system whose isosteric/isoelectric character is considered to be equivalent. And, therefore, they are interchangeable as far as its contribution to biological activity.<sup>5</sup> Here, we report the synthesis and the biological activity of 1-hydroxy-2-pyridone analogue (**2**) of capsaicin.

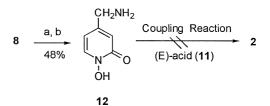
The synthesis of target compound 2 was started with the conversion of pyridine-N-oxide into pyridone compound (Scheme 1). Thus, 4-cyanopyridine-N-oxide 3 was refluxed with acetic anhydride followed by acid hydrolysis with acetic acid to give the pyridone 4. After O-methylation of pyridone,<sup>6</sup> the free amine  $\mathbf{6}$  was obtained from the reduction of the cyano group in 5 with LiAlH<sub>4</sub>. Protection of the free amine with trifluoroacetyl group followed by N-oxidation of the resulting compound 7 with m-chloroperbenzoic acid gave the N-oxide 8, where the trifluoroacetyl group was then deprotected under basic condition to give 4-aminomethyl-2methoxypyridine-1-oxide (9). Coupling reaction of 9 with (E)-8-methyl-6-nonenoic acid (11), which was prepared by the reported method,<sup>7</sup> under DCC/DMAP condition gave 10 in 52% yield. Finally, the methyl group was removed by refluxing with acetyl chloride followed by hydrolysis with acetone-water mixture<sup>8</sup> to give 2. However, it is interesting to note that the coupling reaction of 4-aminomethyl-1hydroxy-2-pyridone (12), which was prepared from 8, with the corresponding acid 11 under various conditions was unsuccessful to obtain 2 (Scheme 2).

The analgesic activity of 2 was determined by the reported





Scheme 1. (a) i)  $Ac_2O$ , reflux, ii) AcOH, reflux, (b)  $Ag_2CO_3$ ,  $CH_3I$ , benzene, (c)  $LiAlH_4$ , ether, (d) TFAA,  $CH_2Cl_2$ , (e) mCPBA,  $CH_2Cl_2$ , (f)  $K_2CO_3$ , MeOH, (g) (E)-8-Methyl-6-nonenoic acid (11), DCC, DMAP,  $CH_2Cl_2$ , (h) i) AcCl, ii)  $H_2O$ , acetone.



Scheme 2. (a) i) AcCl, ii) H<sub>2</sub>O, (b) MeOH : H<sub>2</sub>O : c-HCl (1 : 2 : 2).

method<sup>3a</sup> and observed to be inactive ( $ED_{50} = > 50 \mu mol/kg$ ).<sup>9</sup> It can therefore be concluded that the catechol moiety in capsaicin makes a major contribution to the analgesic activity.

## **Experimental**

Instrument. Melting points were determined on a

Fisher-Johns melting point apparatus and are uncorrected. <sup>1</sup>H NMR and <sup>13</sup>C NMR spectra were recorded on a Varian Gemini 200 spectrometer at 200 MHz and 50 MHz. Chemical shifts were given in relative tetramethylsilane. Infrared spectra were recorded on a Nicolet FT-IR 550 spectrometer. Elemental analyses were performed by Fisons Eager 200 instrument, Italy. Column chromatography was done by using Merck silica gel 60 (230-400 mesh). All reactions were performed under a nitrogen atmosphere.

4-Cyano-2-pyridone (4). A solution of 4-cyanopyridine-N-oxide (50.0 g, 0.140 mol) in acetic anhydride (500 mL) was refluxed for 18 h and then concentrated. The residue was dissolved in ethyl acetate to give black precipitate which was identified with reactant. After filtering the precipitate, the filtrate was concentrated to give a brown residue, which was chromatographed on a silica gel column (ethyl acetate : hexane = 1:1) to give 4-cyano-2-acetoxypyridine (11.5 g, 23%) as a yellow solid. mp 60 °C, IR (KBr) 2243, 1775, 1196 cm<sup>-1</sup>, <sup>1</sup>H NMR (CDCl<sub>3</sub>) δ 2.31 (s, 3H, -CH<sub>3</sub>), 7.33 (s, 1H, 3-py-H), 7.44 (d, 1H, 5-py-H, *J* = 4.8), 8.53 (d, 1H, 6-py-H, J = 4.8), <sup>13</sup>C NMR (CDCl<sub>3</sub>)  $\delta$  20.8, 115.4, 118.7, 123.2, 123.4, 149.7, 158.0, 168.0. A solution of 4cyano-2-acetoxypyridine (8.5 g, 50 mmol) in acetic acid (300 mL) was heated at 100-110 °C for 3 h and then concentrated. The resulting residue was recrystallized from methanol to give 4 (4.4 g, 70%) as a violet solid. mp 235 °C, IR (KBr) 2237, 1676 cm<sup>-1</sup>, <sup>1</sup>H NMR (DMSO-d<sub>6</sub>) δ 6.46 (dd, 1H, 5-py-H, J = 1.0, 6.6 Hz), 6.97 (s, 1H, 3-py-H), 7.65 (dd, 1H, 6-py-H, J = 1.0, 6.6 Hz), 12.25 (br., 1H, -NH), <sup>13</sup>C NMR (DMSO-d<sub>6</sub>) δ 104.8, 116.5, 123.9, 126.0,138.4, 160.8, Anal. Calcd for C<sub>6</sub>H<sub>4</sub>N<sub>2</sub>O: C, 59.99; H, 3.36; N, 23.32. Found: C, 59.78; H, 3.37; N, 23.23.

**4-Cyano-2-methoxypyridine (5).** To a solution of **4** (6.2 g, 50 mmol) in benzene (50 mL) was added silver carbonate (15 g, 54 mmol) and iodomethane (18 mL, 290 mmol). The reaction mixture was stirred for 48 h at room temperature and then concentrated. The brown residue was chromatographed on a silica gel column (ethyl acetate : hexane = 1 : 1) to give **5** (5.0 g, 71%) as a pale yellow solid. mp 64 °C, IR (KBr) 2237, 1041 cm<sup>-1</sup>, <sup>1</sup>H NMR (CDCl<sub>3</sub>) δ 3.97 (s, 3H, -OCH<sub>3</sub>), 7.00 (s, 1H, 3-py-H), 7.08 (dd, 1H, 5-py-H, *J* = 1.1, 5.2 Hz), 8.32 (d, 1H, 6-py-H, *J* = 5.2 Hz), <sup>13</sup>C NMR (CDCl<sub>3</sub>) δ 53.9, 114.1, 116.4, 117.5, 122.3, 148.4, 164.3, Anal. Calcd for C<sub>7</sub>H<sub>6</sub>N<sub>2</sub>O: C, 62.68; H, 4.51; N, 20.88. Found: C, 62.55; H, 4.50; N, 20.64.

**4-Aminomethyl-2-methoxypyridine (6).** To a suspension of LiAlH<sub>4</sub> (1.6 g, 2.0 eq.) in anhydrous ether (200 mL) was slowly added **5** (2.4 g, 18 mmol) in ether (15 mL) at 0 °C. The reaction mixture was stirred for 3 h at room temperature and then quenched with 10% aqueous NaOH solution (10 mL). After the resulting gray precipitate was filtered, the organic solvent was removed. The residue was chromatographed on a silica gel column (ethyl acetate : methanol = 1 : 5) to give **6** (1.7 g, 71%) as a yellow oil. IR (neat) 3367, 3317 cm<sup>-1</sup>, <sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta$  3.84 (s, 2H, -CH<sub>2</sub>), 3.93 (s, 3H, -OCH<sub>3</sub>), 6.70 (s, 1H, 3-py-H), 6.83 (d, 1H, 5-py-H, *J* = 5.2 Hz), 8.08 (d, 1H, 6-py-H, *J* = 5.2 Hz), <sup>13</sup>C NMR (CDCl<sub>3</sub>)

## δ 53.9, 114.1, 116.4, 117.5, 122.3, 148.4, 164.3.

**4-Trifluoroacetylaminomethyl-2-methoxypyridine (7).** To a solution of **6** (1.0 g, 75 mmol) in methylene chloride (100 mL) was slowly added trifluoroacetic anhydride (1.7 mL). After stirring for 1 h at room temperature, the reaction mixture was neutralized with 10% aqueous NaHCO<sub>3</sub> solution and then extracted with methylene chloride ( $3 \times 50$  mL). The organic layer was washed with water ( $2 \times 50$  mL), dried (Na<sub>2</sub>SO<sub>4</sub>), filtered, and concentrated to give **7** (1.37 g, 78%) as a yellow solid. mp 58 °C, IR (KBr) 3310, 1701, 1147 cm<sup>-1</sup>, <sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta$  3.78 (s, 3H, -OCH<sub>3</sub>), 4.34 (d, 2H, -CH<sub>2</sub>, *J* = 6.0 Hz), 6.51 (s, 1H, 3-py-H), 6.67 (d, 1H, 5-py-H, *J* = 5.4 Hz), 7.98 (d, 1H, 6-py-H, *J* = 5.4 Hz), <sup>13</sup>C NMR (CDCl<sub>3</sub>)  $\delta$  41.9, 53.1, 107.9, 118.5, 146.8, 148.3, 158.2, 164.4, Anal. Calcd for C<sub>9</sub>H<sub>9</sub>N<sub>2</sub>O<sub>2</sub>F<sub>3</sub>: C, 46.16; H, 3.87; N, 11.96. Found: C, 46.00; H, 3.87; N, 11.87.

**4-Trifluoroacetylaminomethyl-2-methoxypyridine-N-oxide (8).** To a solution of **7** (0.98 g, 4.2 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (50 mL) was added m-chloroperbenzoic acid (70-75%, 1.8 g). After the resulting mixture was stirred for 12 h at room temperature, the organic solvent was removed. The residue was chromatographed on a silica gel column (ethyl acetate : hexane = 3 : 1) to give **8** (0.75 g, 72%) as a white solid. mp 180 °C, IR (KBr) 3159, 1724, 1211 cm<sup>-1</sup>, <sup>1</sup>H NMR (DMSO-d<sub>6</sub>) δ 3.98 (s, 3H, -OCH<sub>3</sub>), 4.42 (d, 2H, -CH<sub>2</sub>, *J* = 3.1 Hz), 6.93 (d, 1H, 5-py-H, *J* = 6.1 Hz), 7.16 (s, 1H, 3-py-H), 8.23 (d, 1H, 6-py-H, *J* = 6.1 Hz), 10.13 (br., 1H, -NH), <sup>13</sup>C NMR (DMSO-d<sub>6</sub>) δ 41.4, 57.2, 108.1, 116.6, 137.0, 139.3, 156.4, 157.1, 158.0.

**4-Aminomethyl-2-methoxypyridine-N-oxide (9).** To a solution of **8** (0.75 g, 30 mmol) in water (10 mL) and methanol (40 mL) was added potassium carbonate (0.81 g). The reaction mixture was stirred for 5 h at room temperature and then concentrated. The residue was chromatographed on a silica gel column (methanol) to afford **9** (0.15 g, 32%) as a yellow oil. IR (KBr) 3418, 1176 cm<sup>-1</sup>, <sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta$  3.86 (s, 2H, -CH<sub>2</sub>), 4.02 (s, 3H, -OCH<sub>3</sub>), 6.85 (dd, 1H, 5-py-H, *J* = 1.5, 6.7 Hz), 6.95 (s, 1H, 3-py-H), 8.14 (d, 1H, 6-py-H, *J* = 6.7 Hz).

(E)-8-Methyl-6-nonenoic Acid (11). This was prepared by following the procedure reported by H. Kaga *et al.*.<sup>7</sup> Yellow oil. IR (neat) 3409, 1706, 1265 cm<sup>-1</sup>, <sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta$  0.96 (d, 6H, (CH<sub>3</sub>)<sub>2</sub>), 1.42 (q, 2H, C<sub>3</sub>-H), 1.63 (q, 2H, C<sub>4</sub>-H), 2.02 (q, 2H, C<sub>5</sub>-H), 2.32 (m, 1H, C<sub>8</sub>-H), 2.36 (t, 2H, C<sub>2</sub>-H), 5.35-5.38 (m, 2H, CH=CH), <sup>13</sup>C NMR (CDCl<sub>3</sub>)  $\delta$  23.1, 24.2, 26.4, 26.8, 29.2, 34.0, 126.5, 138.0, 180.3.

**1-Oxo-2-methoxypyridyl-8-methyl-6***trans***-nonenamide (10).** The mixture of **9** (0.10 g, 64 mmol), (E)-acid **11** (0.12 g, 69 mmol), DCC (0.12 g, 58 mmol), and a catalytic amount of DMAP in methylene chloride (25 mL) was stirred for 8 h at room temperature. After filtration of the precipitate, the filtrate was concentrated. The resulting residue was chromatographed on a silica gel column (ethyl acetate : methanol = 1 : 1) to give **10** (98 mg, 52%) as a yellow oil. IR (KBr) 3267, 1654, 1265 cm<sup>-1</sup>, <sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta$  0.93 (d, 6H, -(CH<sub>3</sub>)<sub>2</sub>), 1.43 (q, 2H, C<sub>3</sub>-H), 1.64 (q, 2H, C<sub>4</sub>-H), 1.98 (q, 2H, C<sub>5</sub>-H), 2.04 (m, 1H, C<sub>8</sub>-H), 2.26 (t, 2H, C<sub>2</sub>-H), 3.98

Notes

(s, 3H, -OCH<sub>3</sub>), 4.36 (d, 2H, -CH<sub>2</sub>NH-, J = 6.0 Hz), 5.35-5.38 (m, 2H, CH=CH), 6.78 (d, 2H, 3,5-py-H, J = 6.6 Hz), 8.09 (d, 1H, 6-py-H, J = 5.5 Hz), 8.12 (br., 1H, -NH-), <sup>13</sup>C NMR (CDCl<sub>3</sub>)  $\delta$  22.6, 25.2, 29.3. 30.9, 32.2, 36.2, 41.7, 57.2, 107.1, 116.3, 126.3, 138.0, 139.2, 141.9, 157.9, 173.7.

4-Aminomethyl-1-hydroxy-2-pyridone (12). A solution of 8 (150 mg, 6.4 mmol) in acetyl chloride (20 mL) was refluxed for 1 h and then concentrated. The resulting yellow residue dissolved in water (20 mL) was stirred for 14 h at room temperature. After the solvent was evaporated, the crude 1-hydroxy-2-pyridone derivative was obtained as a white solid, which was dissolved in small amount of methanol-water-c-HCl (1:2:2, 20 mL) again and then refluxed for 17 h. After the reaction mixture was concentrated, the yellow precipitate was recrystallized from ethanol-water to give the HCl salt form of 12 (54 mg, 48%) as a yellow solid. mp 230 °C, IR (KBr) 3427, 1650 cm<sup>-1</sup>, <sup>1</sup>H NMR (DMSO-d<sub>6</sub>) δ 3.90 (s, 2H, CH<sub>2</sub>), 6.35 (d, 1H, 3-py-H, J = 5.0 Hz), 6.60 (s, 1H, 5-py-H), 7.94 (d, 1H, 6-py-H, *J* = 7.2 Hz), 8.50 (br., 3H,  $-NH_{3^+}$ ), <sup>13</sup>C NMR (DMSO-d<sub>6</sub>)  $\delta$  104.1, 118.1, 136.0, 144.8, 157.4. Free base was obtained from the following method; after the salt was dissolved in ammonia solution, extraction with methylene chloride followed by general work-up gave the free base, which was used for the coupling reaction immediately

N-Hydroxy-2-oxopyridyl-8-methyl-6-trans-nonenamide (2). A solution of 10 (70 mg, 2.0 mmol) in acetyl chloride (6 mL) was refluxed for 1 h and then neutralized with 10% aqueous NaHCO3 solution. The aqueous solution was extracted with methylene chloride  $(3 \times 10 \text{ mL})$ . The organic layer was washed with water, dried (Na<sub>2</sub>SO<sub>4</sub>), filtered, and concentrated. The resulting yellow residue dissolved in small amount of acetone and water was stirred for 8 h at room temperature. The precipitate was filtered and then dried to give 2 (50 mg, 71%) as a slightly yellow solid. mp 107 °C, IR (KBr) 3410, 3279, 1639 cm<sup>-1</sup>, <sup>1</sup>H NMR (CDCl<sub>3</sub>) δ 0.96 (d, 6H, (CH<sub>3</sub>)<sub>2</sub>), 1.35 (q, 2H, C<sub>3</sub>-H), 1.64 (q, 2H, C<sub>4</sub>-H), 1.98 (q, 2H, C<sub>5</sub>-H), 2.15 (m, 1H, C<sub>8</sub>-H), 2.27 (t, 2H, C<sub>2</sub>-H), 4.24 (s, 2H, -CH<sub>2</sub>NH-), 5.34 (m, 2H, CH=CH), 6.21 (s, 1H, 5-py-H), 6.40 (s, 1H, 3-py-H), 7.00 (br., 1H, -NH-), 7.64 (d, 1H, 6-py-H, J = 2.1 Hz), <sup>13</sup>C-NMR (CDCl<sub>3</sub>)  $\delta$  19.5, 20.8,

21.7, 22.6, 24.5, 30.2, 30.4, 92.9, 107.1, 115.8, 123.3, 124.5, 126.4, 141.8, 176.8.

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## References

- Szolcsanyi, J., In Handbook of Experimental Pharmacology. Pyretics and Antipyretics; Milton, A. S., Ed.; Springer: New York, 1982; pp 437-478.
- 2. Fitzgerald, M.; Capsaicin and Sensory Neuron- A Review, *Pain* **1983**, *15*, 109.
- 3. (a) Walpole, C. S. J.; Wrigglesworth, R.; Bevan, S.; Campbell, E. A.; Dray, A.; James, I. F.; Perkins, M. N.; Reid, D. J.; Winter, J. J. Med. Chem. 1993, 36, 2362. (b) Walpole, C. S. J.; Wrigglesworth, R.; Bevan, S.; Campbell, E. A.; Dray, A.; James, I. F.; Perkins, M. N.; Reid, D. J.; Winter, J. J. Med. Chem. 1993, 36, 2373. (c) Walpole, C. S. J.; Wrigglesworth, R.; Bevan, S.; Campbell, E. A.; Dray, A.; James, I. F.; Perkins, M. N.; Reid, D. J.; Winter, J. J. Med. Chem. 1993, 36, 2381. (d) Wrigglesworth, R.; Walpole, C. S. J.; Bevan, S.; Campbell, E. A.; Dray, A.; James, I. F.; Perkins, M. N.; Reid, D. J.; Winter, J. J. Med. Chem. 1996, 39, 4942. (e) Park, N-S.; Choi, J-K.; Hong, M-S.; Lee, J. C.; Choi, S. W.; Lee, B-Y. Korean J. Med. Chem. 1993, 3, 142. (f) Back. G. H.; Jung, Y. S.; Cho. S. J.; Seong, C. M.; Park, N-S. Arch. Pharm. Res. 1997, 20, 659.
- Yoon, S-H.; Bodor, N. S.; Simpkins, J. W. Drug Design and Discovery 1993, 10, 35.
- Friedman, H. L. Influence of isosteric replacements upon biological activity. Symposium on Chemical-Biological correlation. National Academy of Science-National Research Council, Publication no. 206, Washington D. C. 1951; p 295.
- Chung, m. M.; Tiecklmann, H. J. Org. Chem. 1970, 35, 2517.
- Kaga, H.; Miura, M.; Orito, K. J. Org. Chem. 1989, 54, 2477.
- 8. Paquette, L. A. J. Am. Chem. Soc. 1965, 87, 5186.
- Mouse tail flick test was performed at Center for Drug Discovery, University of Florida, U.S.A. according to the procedure described in reference 3a.