

A New Efficient Method for the Synthesis of *L*-Galactose<sup>†</sup>

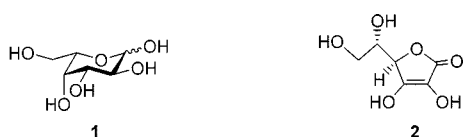
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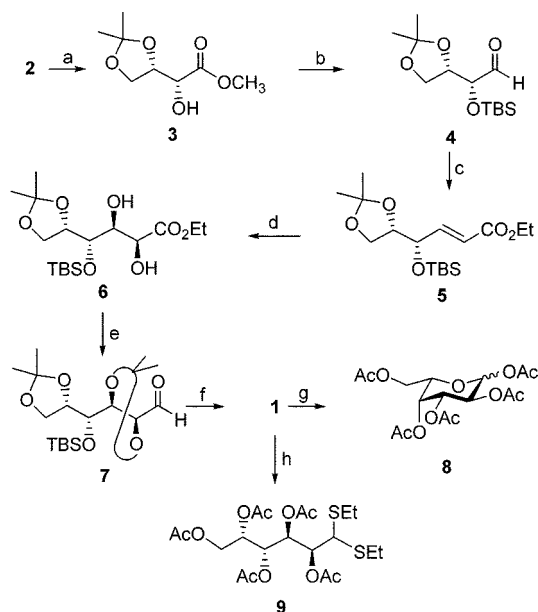
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There is a growing need for the synthesis of non-natural *L*-sugars and naturally occurring rare *L*-sugars because of the medicinal potential of *L*-carbohydrates and related nucleosides due to their potent biological activity and lower toxicity compared to their *D*-counterparts.<sup>1</sup> *L*-Sugars are also used as the building block for the synthesis of *L*-oligonucleotides and enantio-DNA (DNA having *L*-sugar), which are valuable tools for studying protein-DNA interactions and are promising antisense agents.<sup>2</sup> Although certain *L*-sugars such as *L*-fucose, *L*-rhamnose, and *L*-arabinose are quite abundant in nature, *L*-galactose is a rare sugar and occurs as a minor component in agar-agar, chagual gum, red algae, flaxseed mucilage and a snail galactan.<sup>3</sup> There have been reports for the synthesis of *L*-galactose: (i) a synthesis by reduction of *L*-galactono-1,4-lactone,<sup>4</sup> (ii) a method based on the repeated asymmetric epoxidation starting from achiral 2-butene-1,4-diol,<sup>5</sup> (iii) a synthesis employing the Pummerer rearrangement starting from 6-*S*-phenyl-6-thio-*D*-galactose,<sup>6</sup> and (iv) an enzymatic synthesis by galactose oxidase-catalyzed oxidation of galactitol.<sup>7</sup> These methods have some limitations such as the lengthy synthesis, the carefully controlled reaction in certain steps, and/or the low yield of the product. Herein we report an efficient new method for the synthesis of *L*-galactose (**1**) starting from readily available inexpensive *L*-ascorbic acid (**2**).



The synthesis commenced with transformation of *L*-ascorbic acid (**2**) into the methyl threonate **3** in 74% yield by the known procedure.<sup>8</sup> The hydroxyl group of the compound **3** was protected with *t*-butyldimethylsilyl (TBS) chloride (Scheme 1). The resulting TBS ether was subjected to reduction with DIBAL-H at 78 °C to give the aldehyde **4** in 87% yield. Wittig reaction of the aldehyde **4** with Ph<sub>3</sub>P = CHCO<sub>2</sub>Et in the presence of a catalytic amount of benzoic acid provided the (*E*)- $\alpha,\beta$ -unsaturated ester **5** in 93% yield along with a small amount of (*Z*)-isomer (*E*/*Z* = 20 : 1). Dihydroxylation<sup>9</sup> of the compound **5** utilizing AD-mix- $\beta$  in the presence of MeSO<sub>2</sub>NH<sub>2</sub> in *t*-BuOH/H<sub>2</sub>O afforded exclusively the diol **6** in 93% yield. Protection of the diol **6** with 2,2-dimethoxypropane followed by reduction of the



**Scheme 1. Reagents and conditions:** (a) see reference 8, 74% in 3 steps; (b) (i) TBSCl, imidazole, DMF, rt, 12 h, 98%; (ii) DIBAL-H, CH<sub>2</sub>Cl<sub>2</sub>, -78 °C, 1 h, 87%; (c) Ph<sub>3</sub>PCHCO<sub>2</sub>Et, benzoic acid (cat.), CH<sub>2</sub>Cl<sub>2</sub>, rt, 4 h, 93%; (d) AD-mix- $\beta$ , MeSO<sub>2</sub>NH<sub>2</sub>, *t*-BuOH-H<sub>2</sub>O, rt, 30 min, then **5**, 0 °C, 12 h, 93%; (e) (i) 2,2-dimethoxypropane, TsOH (cat.), acetone, 4 h, 96%; (ii) DIBAL-H, CH<sub>2</sub>Cl<sub>2</sub>, -78 °C, 1 h, 89%; (f) *c*-HCl, CH<sub>3</sub>CN-H<sub>2</sub>O, rt, 1 h; (g) Ac<sub>2</sub>O, DMAP (cat.), pyridine, 0 °C to rt, 5 h, 80% in 2 steps from **7**; (h) (i) EtSH, *c*-HCl, rt, 10 min; (ii) Ac<sub>2</sub>O, DMAP (cat.), pyridine 0 °C to rt, 4 h, 91% in 3 steps from **7**.

resultant di-*O*-isopropylidene ester with DIBAL-H at 78 °C gave the protected *L*-galactose **7**.<sup>10</sup> Hydrolysis of the purified **7** with *c*-HCl in acetonitrile provided *L*-galactose (**1**), of which acetylation with acetic anhydride in the presence of a catalytic amount of DMAP in pyridine gave the *L*-galactose pentaacetate **8** in 80% yield in two steps. The crude aldehyde **7** could be used without purification for the subsequent hydrolysis and acetylation steps. <sup>1</sup>H and <sup>13</sup>C NMR spectra of the compound **8** were identical with those of *D*-galactose pentaacetate, which we prepared from *D*-galactose. For the purpose of further identification, *L*-galactose (**1**) was treated with EtSH in the presence of *c*-HCl to afford the acyclic *L*-galactose dithioacetal as white solid, of which acetylation with acetic anhydride gave the pentaacetyl-*L*-galactose dithioacetal **9** {[ $\alpha$ ]<sub>D</sub> -10.7 (c 3.4, CHCl<sub>3</sub>)}. <sup>1</sup>H and <sup>13</sup>C NMR spectra of the compound **9** was identical with those of its enantiomer, pentaacetyl-*D*-galactose dithioacetal {[ $\alpha$ ]<sub>D</sub> +10.5 (c 3.4, CHCl<sub>3</sub>) (lit<sup>11</sup>: [ $\alpha$ ]<sub>D</sub> +9.8, CHCl<sub>3</sub>) (lit<sup>12</sup>: [ $\alpha$ ]<sub>D</sub> +11.31, c

<sup>†</sup>This paper is dedicated to the late Professor Sang Chul Shim.

2.2, CHCl<sub>3</sub>), which we prepared from *D*-galactose. Thus, the conversion of *L*-ascorbic acid to the *L*-galactose pentaacetate **8** was accomplished in 37% overall yield.

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- Compound **7**: <sup>1</sup>H NMR (250 MHz, CDCl<sub>3</sub>) δ 0.12 (s, 6H), 0.92 (s, 9H), 1.31 (s, 3H), 1.33 (s, 3H), 1.41 (s, 3H), 1.50 (s, 3H), 3.67-3.75 (m, 1H), 3.89-3.93 (m, 1H), 3.96-4.08 (m, 3H), 4.57 (dd, *J* = 6.5, 1.1 Hz, 1H), 9.81 (d, *J* = 1.1 Hz, 1H); <sup>13</sup>C NMR (63 MHz, CDCl<sub>3</sub>) δ -4.11, -3.96, 18.44, 25.62, 26.07, 26.44, 26.48, 65.93, 73.37, 77.86, 78.31, 80.38, 109.12, 110.62, 201.46.
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