

15. Chalifa, V.; Mohn, H.; Liscovitch, M. *J. Biol. Chem.* **1990**, 265, 17512.
16. Okamura, S.; Yamashita, S. *J. Biol. Chem.* **1994**, 269, 31207.
17. Massenber, D.; Han, J.-S.; Liyange, M.; Patton, W. A.; Rhee, S. G.; Moss, J.; Vaughan, M. *Proc. Natl. Acad. Sci. U. S. A.* **1994**, 91, 11718.
18. Vinggaard, A. M.; Hansen, H. S. *Biochim. Biophys.*

Acta **1995**, 1258, 169.

19. Choi, S.-W.; Choi, M. *J. Korean Chem. Soc.* **1997**, 41, 672.
20. Jung, K.; Koh, E.; Choi, M. *Bull. Korean Chem. Soc.* **1989**, 10, 585.
21. Kim, M. J.; Kim, B.-S.; Lee, S. Y.; Sohn, J. W.; Kim, C. Y.; Choi, M.-S.; Choi, M. *Bull. Korean Chem. Soc.* **1997**, 18, 1204.

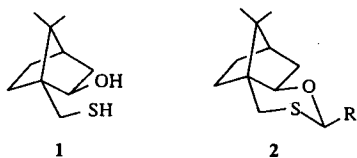
A Convenient Preparation of (1S)-(+)-10-Mercaptoisoborneol from (1S)-(+)-10-Camphorsulfonyl Chloride with High Diastereoselectivity

Kwang-Youn Ko* and Kwang-Il Kim

Department of Chemistry, Ajou University, Suwon 442-749, Korea

Received November 4, 1997

Chiral auxiliaries and catalysts derived from (+)-10-mercaptoisoborneol (*exo*-2-hydroxy-10-mercaptobornane, **1**) are used in various asymmetric syntheses.¹⁻⁵ Recently, 1,3-oxathianes **2** prepared from **1** have been used as highly effective chiral catalysts for the asymmetric epoxidation of aldehydes.⁶



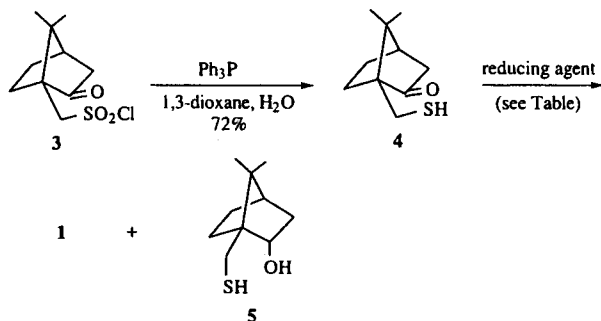
According to the literature,¹ thiol **1** was obtained by reduction of (+)-10-camphorsulfonyl chloride (**3**) with excess lithium aluminum hydride (chloride:hydride=1:4 molar ratio) in ether, followed by chromatographic separation of a 4:1 mixture of **1** and 10-mercaptoborneol (**5**). Because a large excess of pyrophoric reducing agent is employed, handling of this reagent and acidic workup need a careful manipulation. Also, the stereoselectivity in the ketone reduction is not so high. In this paper, we wish to describe a highly stereoselective method of obtaining **1**, which does not use

LiAlH_4 , rendering this method applicable to a large scale preparation, as shown in Scheme 1.

First, sulfonyl chloride **3** was converted to (+)-camphor-10-thiol (**4**) with triphenylphosphine in dioxane-water co-solvent.⁷ Thiol **4** could be easily separated from the reaction mixture by extraction with 5 M NaOH solution followed by acidification and extraction of the resulting thiol with hexanes. Sublimation of the crude product at reduced pressure gave the crystalline ketone **4** in 72% yield. Next, we studied the diastereoselective reduction of ketone **4** (Table 1). Reduction with LiAlH_4 or *i*- Bu_2AlH gave *exo*-isomer **1** in high selectivity (>95% de). Similarly, NaBH_4 in EtOH showed a high *exo*-selectivity.⁸ After reduction, removal of the minor *endo*-isomer **5** (polar) by silica gel column chromatography (hexanes:ethyl acetate=20:1) gave the diastereomerically pure *exo*-thiol **1**. This thiol is slowly oxidized to the disulfide when exposed to the air. Therefore, it should be kept under an inert atmosphere in a cold place.

Experimental Part

The ^1H and ^{13}C NMR spectra were recorded with a Vari-



Scheme 1.

Table 1. Reduction of ketone **4** with reducing agents^a

Reducing Agent	Solvent	Temp. (°C)	Time	1/5 ^b
LiAlH_4	THF	-78	3 h	≥95/5
LiAlH_4	THF	0	3 h	≥95/5
LiAlH_4	ether	-78	3 h	92/8
LiAlH_4	ether	0	3 h	≥95/5
NaBH_4	ethanol	rt	2 day	95/5
<i>i</i> - Bu_2AlH	toluene	-78	3 h	95/5

^aChemical yield was >95% in all cases. ^bThe ratio was determined by the integration of two sets of peaks around 3.97 ppm (*exo*) and 4.35 ppm (*endo*) in ^1H NMR spectrum. It should be noted that the ratio determined by the ^1H NMR method can have an error of ±5%.

an Gemini 200 spectrometer in CDCl_3 using TMS as an internal standard. The FT-IR spectra were measured on a Nicolet 500 spectrometer as KBr disks. Optical rotations were measured on a Jasco DIP-370 digital polarimeter.

(+)-Camphor-10-thiol (**4**). A solution of (+)-10-camphorsulfonyl chloride (**3**, 18.0 g, 71.8 mmol) in a mixture of 240 mL of dioxane and 60 mL of water was treated with triphenylphosphine (75.3 g, 287 mmol). The clear mixture was stirred for 2 days at room temperature, and then refluxed for 1 h. The reaction mixture was concentrated in vacuo. The resulting syrupy residue was extracted with hot hexanes (200 mL \times 3). As the hexanes solution cooled to room temperature, a white precipitate (phosphine oxide) was formed, which was discarded. The hexanes solution was extracted with 5 M NaOH solution (100 mL \times 5) and the combined NaOH extract cooled in an ice bath was acidified by careful addition of concentrated HCl (250 mL). The thiol was separated as a white solid, which was extracted with ethyl acetate (200 mL \times 3). The organic solution was dried (Na_2SO_4) and concentrated in vacuo to give a white solid. Finally, purification by vacuum sublimation (125-135 $^\circ\text{C}$, 0.05 mmHg) yielded 9.47 g (72%) of the thiol **4** as a colorless crystal, mp 65-66 $^\circ\text{C}$ (lit.⁷ 65-67 $^\circ\text{C}$); $[\alpha]_{\text{D}}^{20} = +5.2$ ($c = 1.02$, CHCl_3); ^1H NMR (CDCl_3) δ 4.00-3.98 (m, 1H), 2.79 (dd, 1H, $J = 9.5$ and 13 Hz), 2.58 (dd, $J = 5.3$ and 13 Hz), 2.15 (bs, 1H), 1.28 (dd, 1H, $J = 9.5$ and 5.3 Hz), 1.05 (s, 3H), 0.83 (s, 3H), and others; ^{13}C NMR (CDCl_3) δ 217.2, 60.3, 47.5, 43.4, 42.9, 26.7, 26.3, 21.0, 20.0, 19.5; IR (KBr) cm^{-1} 1731.

(1S)-(+)-10-Mercaptoisoborneol (**1**). To a solution of NaBH_4 (2.06 g, 54.2 mmol) in EtOH (100 mL), cooled in an ice bath was added a solution of ketone **4** (5.00 g, 27.1 mmol) in EtOH (20 mL) over 10 min under a nitrogen atmosphere. The whole mixture was stirred for 2 days. Then, the excess NaBH_4 was destroyed with dilute HCl solution. The product was extracted with EtOAc (100 mL \times 2). The combined extract was washed with brine, dried (Na_2SO_4) and concentrated in vacuo. Finally, column chromatography of the residue (eluent; hexanes:EtOAc=20:1) on silica gel gave 4.73 g (93% yield) of the product as a solid, mp 73-74 $^\circ\text{C}$ (lit.¹ 76-78 $^\circ\text{C}$; lit.³ 7 $^\circ\text{C}$); $[\alpha]_{\text{D}}^{20} = -56.0$ ($c = 1.15$, CHCl_3)

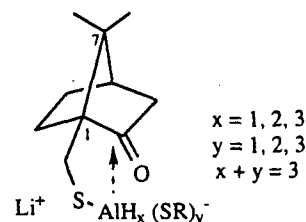


Figure 1.

(lit.¹ $[\alpha]_{\text{D}}^{24} = -55.4$; lit.³ $[\alpha]_{\text{D}}^{24} = -57.44$ ($c = 10$, CHCl_3); ^1H NMR (CDCl_3) δ 3.97 (apparent t, 1H, $J = 4.7$ Hz), 2.79 (dd, 1H, $J = 9.5$, 13 Hz), 2.56 (dd, 1H, $J = 5.4$, 13 Hz), 1.28 (dd, 1H, $J = 9.5$, 5.4 Hz), 1.05 (s, 3H), 0.83 (s, 3H), and others; ^{13}C NMR (CDCl_3) δ 76.4, 52.9, 47.4, 45.7, 39.4, 30.3, 26.8, 23.7, 20.5, 19.9; IR (KBr) cm^{-1} 3467, 2950, 1393, 1373, 1071, 1033.

Acknowledgments. This study is supported by the academic research fund of Korean Ministry of Education (BSRI-96-3149, BSRI-97-3449).

References

1. Eliel, E. L.; Frazee, W. J. *J. Org. Chem.* **1979**, *44*, 3598.
2. Isobe, M.; Obeyama, J.; Funabashi, Y.; Goto, T. *Tetrahedron Lett.* **1988**, *29*, 4773.
3. De Lucchi, O.; Luccini, V.; Marchioro, C.; Valle, G.; Modena, G. *J. Org. Chem.* **1986**, *51*, 1457.
4. Annunziata, O.; Cinquini, M.; Cozzi, F.; Farina, S.; Montanari, V. *Tetrahedron* **1987**, *43*, 1013.
5. Eschler, B. E.; Haynes, R. K.; Kremmydas, S.; Ridley, D. *D. J. Chem. Soc., Chem. Commun.* **1988**, 137.
6. Aggarwal, V. K.; Ford, J. G.; Thompson, A.; Jones, R. V. H.; Standen, M. C. H. *J. Am. Chem. Soc.* **1996**, *118*, 7004.
7. Oae, S.; Togo, H. *Bull. Chem. Soc. Jpn.* **1983**, *56*, 3802.
8. The high selectivity may be ascribed to the model above (Figure 1) where the carbon-sulfur bond is *anti* to the C1-C7 bond and the hydrogen atom is transferred intramolecularly to the less hindered si face of the carbonyl bond.

Highly Overlapping ^1H NMR Signal Assignments of 12,13-Diepimeric Coenzyme F430 by the Compensated ROESY Experiment

Hoshik Won* and Michael F. Summers†

Department of Chemistry, Hanyang University, Ansan 425-791, Korea

†Department of Chemistry and Biochemistry, University of Maryland Baltimore County, Baltimore, MD 21228, U.S.A.

Received November 5, 1997

Coenzyme F430 is a nickel(II)-containing cofactor of the methyl coenzyme M reductase (Component C which was found in the cells of methanogenic bacteria) that is involved in the bio-catalytic reduction of methyl coenzyme M (2-methylthioethanesulfonic acid, $\text{CH}_3\text{-S-CoM}$).^{1,2} Coenzyme

F430 is known to be mediated in the reductive demethylation of methyl coenzyme M, using reducing equivalents from 7-mercaptoheptanoylthreonine phosphate (HS-HTP). The products of this reaction are methane and the heterodisulfide of methyl coenzyme M and HS-HTP (CoM-S-S-HTP).³⁻⁵