Thermostatic regulation of blood samples in blood gas analysers: results and improved method applied to 11 different models

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Introduction

As part of a general assessment of blood gas analysers [1] the thermostatic regulation of blood samples in the machines' sample chambers was studied with the aid of a thermal probe placed on one of the electrodes. The experiment was necessary because the analysers' readings assume a temperature of 37° C; in principle the instruments should maintain this temperature to within 0.1°C.

The influence of temperature varies with each parameter being measured (for example pH, percentage oxygen $[pO_2]$ and percentage carbon dioxide $[pCO_2]$) and according to the nature of the solution being analysed [2]. For protein solutions, and especially for blood samples, the average variations are: for pH, 0011 units per degree; for pCO_2 , 4%/°C, and for pO_2 , 7%/°C. In addition, cooling of the electrodes modifies these variations: for pO_2 this may vary by as much as 10%/1°C; for pCO_2 the effects of temperature are in part compensatory, reducing the variations to -2% error/1°C [3, 4].

Aqueous buffers are less sensitive to temperature; they are not therefore suitable for thermostatic control [5].

The measuring electrode in each instrument was modified. The sensing part of the pO_2 or the pCO_2 was replaced by the heat-sensitive end of a thermocouple.

The aim of this investigation was to examine the temperature of the sample itself within the sample chamber, without disturbing the normal flushing, calibrating and measuring processes which are very elaborate in some 'automatic' instruments.

Materials and methods

pH blood gas analyser

A wide range of instruments, 11 different models, produced by five international manufacturers (Corning, BP 36, 11 Chemin de Ronde, F 78110 Le Vesinet, France; Technicon, Route Nationale No. 1, F 95330 Domont, France; Instrument Laboratory—Delhomme, 32 avenue St Mandé, F 75562 Paris Cedex 12; Radiometer—Jarre Jacquin, 30 rue St Augustin, F 75002 Paris; and AVL, Place de l'Europe, F 94019 Creteil, France) were tested. See table 1 for a list of the machines and the results obtained. For each model (except one) two instruments were tested to ensure the validity of the results.

Thermocouple

The thermocouple used was a Constanant which has a diameter of 1 mm, a very short response delay of 3 s, and a theoretical sensitivity of 4 MV/°C. Thus 0.01°C can be estimated ensuring a precision 0.1°C.

The thermocouple was fixed on each instrument's jacket in place of the pCO_2 electrode. The thermocouple was passed through a silicon rubber plug (cut to size), and pushed firmly to the bottom of the jacket in order to keep the system watertight. A large diameter trocar (3 mm) was used to pass the thermocouple through this stopper. Once the trocar is removed, the thermocouple remains held in place by the elasticity of the stopper (see figures 1 and 2).



Figure 1. Parts required for assembling the thermocouple.

Each electrode assembly fitted with the thermocouple was then calibrated in a water bath with a circulating system, at three or four different temperatures ranging from 36° C to 38° C.

The same high-precision thermometer (manufactured by Walter-Kessler, Z.A. Courtaboeuf, F 91403 Orsay, France), graduated to 0.02°C, was used throughout the investigation. A calibration curve was drawn for each instrument (figure 3). Zero was obtained by plunging the two ends of the thermocouple into a thermos flask filled with distilled water and ice. The iced water served throughout each experiment as a fixed temperature of reference.



Figure 2. Thermocouple fixed on an instrument's jacket. Note the fineness of the sensitive part of the thermocouple.



Figure 3. Calibration of the thermocouple fixed on the instruments' jackets.

Recorder

The recorder used was a Kipp and Zonnen micrograph BDN°5. Temperatures were recorded during the instrument calibration with gas; during three successive measurements of one blood sample previously maintained at 25°C, and during the three successive measurements of the same blood sample previously maintained at $+4^{\circ}$ C. In each case the manufacturer's operating instructions were followed and the timing programmes of the instruments were taken into account. The thermocouple system indicates the temperature variations in the sample chamber when the gas is passed through, although the values obtained, due to the gas, are not necessarily exact. The continuous recordings of temperatures from within the sample chamber are illustrated in figure 4.

Results

The results of the study are shown in table 1; the instruments are listed there according to their degree of automation. The notes below relate to the table.

(a) In this instrument the galvanometer indicating temperature reverts to 37°C 30s before complete thermal equilibrium of the sample.



Figure 4. Examples of recorded temperatures (1 square = approximately 0.13°C). Figure 4(a) is from an instrument without a pre-heater (speed of paper was 10 mm/min.) and figure 4(b) is from an instrument with a pre-heater (speed of paper was 5 mm/min.); a = rinsing, b = entry of sample, c = final temperature.

- (b) The instrument confirms its results after about 45s although the thermal balance of the sample is reached after 2 min. 10s at 37.12°C.
- (c) After rinsing with the non-thermostated solutions the temperature of the measuring chamber is slow in returning to its exact level (3 min.). In addition it was noted with this instrument that too great a speed of gas flow significantly lowered the temperature of the chamber (the calibration gases do not pass through the preheater). The flow rates advised by the manufacturer should be followed.
- (d) See note (b).
- (e) This instrument was adjusted by the manufacturer just before testing: this could explain the quality of the results.
- (f) The thermal equilibrium of the sample is reached after about $2 \min$. 30 s at $36 \cdot 85^{\circ}$ C. The instrument thus confirms its results before thermal equilibration is reached, hence the difference in values observed.
- (g) After reassembly of the instrument the test was not carried out until the temperature warning light was extinguished.

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		Blood stored at 25°C			Blood stored at +4°C				
Instrument		Values of the three readings in °C			Valu re	ies of the t adings in ^c	- Isothermic time lag	Remarks	
Corning 161	(1)	36·94	36·94	36·94	36·94	36·90	36·90	1 min. 20s	a
	(2)	36·85	36·83	36·87	36·81	36·78	36·80	1 min. 30s	a
BMS MK ₂	(1)	36·88	36·76	36·82	36·80	36·80	36·86	3 min.	
Radiometer	(2)	36·62	36·62	36·62	36·62	36·62	36·62	3 min.	
IL 213		36.90	36.85	36.85	36.86	36.90	36.90	3 min.	
Corning 166	(1)	37·00	36·85	36·85	36·85	36·85	36·76	2 min. 20 s	a
	(2)	37·02	37·04	37·02	37·05	37·05	37·05	2 min. 20 s	a
AVL 938		36.60	35.0		36.66	35.02		2 min. 10 s	b
IL 413	(1)	36·62	36·68	36·68	36·68	36·68	36·68	5 s	с
	(2)	36·72	36·70	36·72	36·50	36·56	36·56	5 s	с
IL 613	(1)	36·75	36·60	36·60	36·65	36·60	36·60	5s	c
	(2)	36·69	36·63	36·60	36·64	36·60	36·70	5s	c
AVL 940	(1)	36·46	36·53	36·60	36·46	36·50	36·60	1 min. 30 s	d
	(2)	(37·05)	(37·05)	(36·94)	(37·05)	(37·05)	(36·96)	1 min. 30 s	e
Corning 175	(1)	36·98	36·98	37·04	37·04	37·16	37·10	50 s	a
	(2)	37·10	37·12	37·12	37·12	37·10	37·12	1 min.	a
Technicon BG I	I (1)	36·60	36·68	36·68	36·66	36·48	36·75	2 min. 30 s	f
	(2)	36·44	36·28	36·55	36·50	36·48	36·56	2 min. 30 s	f
ABL ₂	(1)	36·24	36·30	36·30	36·30	36·30	36·30	10 s	$g \ g$
Radiometer	(2)	36·48	36·46	36·46	36·46	36·46	36·44	10 s	

Table 1.	Temperature	of	`blood	samples	in	the	measurina	chamber.
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The table shows temperatures of samples when the operator, or automatic instrument, confirms the result, but not necessarily the temperature of the sample at calorific balance.

Accuracy

The lowest temperature measured was 36.24° C and the highest 37.16° C. One company, Corning, stands out as ensuring a high quality of sample thermoregulation on the whole range of their instruments, irrespective of the level of automation. All other models show greater or lesser inexactitude, erring always by default.

Repeatability

Table 1 demonstrates that, on the whole, the repeatability of temperature is very good for a given instrument.

Influence of sample temperature

It can equally be seen that the initial temperature of the sample does not affect its final temperature in the sample chamber. This holds good whether the instrument has a pre-heater or not. This fact means that if measuring has to be deferred (due to work-loads in the laboratory or any other reason) the blood samples can be refrigerated (at $+4^{\circ}$ C).

Comments

It was noticed in the course of these investigations that too high a flow rate of gas in the sample chamber during calibration cooled the chamber (from between 1 and 1.5° C) and this invalidates calibration values.

In some instruments (AVL particularly), the copious flow of cleaning fluids which are not thermostatically controlled leads to significant cooling in the sample chamber, which then takes about 5 min. to regain its correct temperature this reduces the analytical throughput of samples.

For instruments which have no fixed time input the steady state of sample temperature may be reached if the response time of electrodes is slow. On the other hand, when the response time is fast, the results are read before the steady state.

When pre-heaters are provided with the instrument the blood sample enters the sample chamber at a temperature very near to the steady state.

Conclusion

Twenty instruments—11 models from five manufacturers were tested; it was concluded that only one company offers instruments which conform technically with the documentation supplied.

The other companies' instruments fail in accuracy but good repeatability of sample thermostatization is obtained; therefore it should be possible to overcome this problem by correctly adjusting the instruments.

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