

# An automated determination of beta-glucuronidase activity in human serum with the Abbot VP bichromatic analyzer

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## Introduction

The determination of serum beta-glucuronidase (b-D-glucuronide glucuronohydrolase, EC 3.2.1.31) is of clinical interest because it can be used in the diagnosis of several pathological conditions. Serum levels of beta-glucuronidase are increased in patients with such conditions as neoplasms [1], diabetes mellitus [2 and 3], in pregnant women and in gestational diabetes [4], atherosclerotic disease [5], coronary artery disease [6], and Gaucher's disease [7]. However, it is practically absent in sera of patients with mucopolisaccharidosis [8].

When a glycoside of a b-D-glucosiduronic acid is used as substrate, beta-glucuronidase catalyses the transfer of b-D-glucosiduronic acid to an appropriate acceptor such as water (hydrolysis reaction), alcohols and glycols.

Fluorimetric and colorimetric measurement, based on the hydrolysis reaction, can be carried out for the determination of beta-glucuronidase activity [9]. In the colorimetric methods, three substrates are widely used—the glucuronides of phenolphthalein [10], 4-nitrophenol [11], and 8-hydroxyquinoline [12], which upon enzymatic action liberate the corresponding aglycone. These are coloured substances in alkaline mediums. 4-nitrophenol-b-D-glucuronide was chosen as the substrate for the authors' beta-glucuronidase assay because 4-nitrophenol is the final product in at least 20 different enzymatic determinations [13].

In this paper, an optimized method [14] for determining the enzymatic activity of beta-glucuronidase has been adapted for the Abbott VP Bichromatic Analyzer. In the reference method, the incubation time was 1 h, the substrate concentration was 8mM in the assay and the CVs calculated were: within-run CV = 8.86  $\bar{x}$  = 1.05; CV = 4.99,  $\bar{x}$  = 4.47; between-day CV = 7.75,  $\bar{x}$  = 2.85.

## Materials and methods

### Reagents

Acetic acid, sodium acetate, and sodium hydroxide, all analytical grade reagents from Merck, Darmstadt, FR Germany. 4-nitrophenyl-b-D-glucuronide, 4-nitrophenol, b-glucuronidase from bovine liver and glycine from Sigma Chemical Co., St. Louis, Missouri, USA.

### Apparatus

UV-Vis spectrophotometer, Minikem with printer (Coulter Electronics, Inc., Hialech, Florida, USA). pH meter 28 (Radiometer Copenhagen, Denmark). Analytical balance H 10 W and precision balance P 1210 from Mettler Instrumente AG, Switzerland. Abbott VP Bichromatic Analyzer (Abbott Laboratories, Dallas, Texas, USA).

### Samples

The blood samples used were collected from healthy volunteers, blood donors, and pregnant women attending the Hospital 'San Agustín'. The sera were stored in a refrigerator at 4–8 °C until the determination of enzymatic activity was carried out. The use of heparinized plasma was avoided because heparin has been described as inhibiting beta-glucuronidase activity [15]. The use of haemolysed sera was also avoided.

Two pools of sera were made, one normal and the other pathological, which were used as controls; these were stored at –20 °C. Frozen samples may be analysed if mixed thoroughly after thawing.

### Method

Before analysis the instrument is set as follows:

TEST NAME	USER TEST
TEMPERATURE	37 °C
FILTER VALUE	415/450
UNITS OF MEASUREMENT	IU/1
DILUTION RATIO SETTING	1 : 11
AUXILIARY DISPENSER?	YES
AUXILIARY DISPENSE STATION	11
AIR MIX?	NO
AUXILIARY DISPENSE VOLUME	150.34
FRR?	YES
REACTION DIRECTION	UP
STANDARDS?	NO
REAGENT BLANK?	YES
ASSAY FACTOR	696.1
TEST TYPE?	END POINT
AUXILIARY DISPENSE	
REVOLUTION	8
BGN PRT REVOLUTION	9
PRT REVOLUTION	1
INITIAL REAGENT AD	0.2
UP LIMIT?	YES
MAX ABSORBANCE LIMIT	1.5

In the test-tube, the main syringe deposits the substrate p-nitrophenyl-b-D-glucuronide (4 mM in the assay,

dissolved in the buffer HAc/Ac 0.1 M pH = 4.0, with 25  $\mu$ l of the sample). The auxiliary dispenser is filled with buffer glycine-NaOH 0.5 M pH = 10.6. Water is placed in position No.1 of the carousel, and the different samples are placed in positions 2 through 31. In the first assays, water was deposited in the 00 position of the carousel, in order to ensure that the substrate was not undergoing hydrolysis.

The assay factor is calculated using standards containing 1.0 mM 4-nitrophenol which are processed as a normal samples. Results are thus obtained directly in IU/1.

## Experimental results

### Analytical variables

#### Linearity

As shown in figure 1, the method is linear over a wide range of activity values, since those up to 110 IU/1 can be measured without dilution. Because these high activities are rarely found in human sera, purified beta-glucuronidase from bovine liver was used for this experiment.

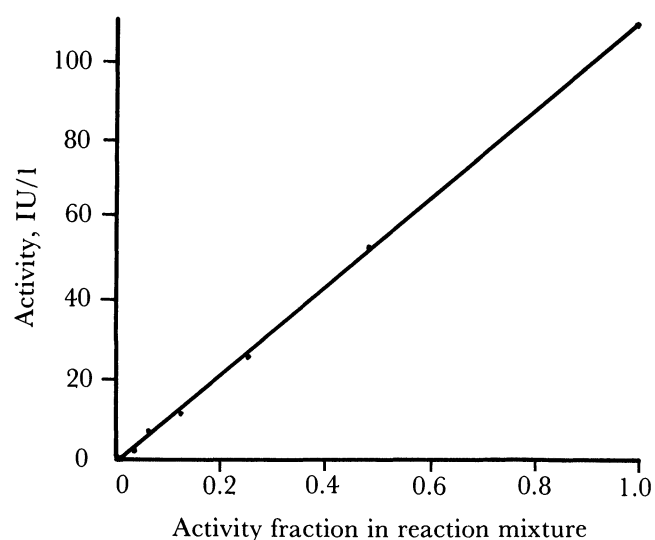


Figure 1. Linearity of the method of assay. A sample of beta-glucuronidase from bovine liver was serially diluted with isotonic saline solution to prepare different dilutions, and the activity of the enzyme determined in all of them.

#### Precision

Table 1 shows the precise data from the assay. Samples with normal and high activities were assayed to estimate the within-run variation. To estimate the between-day variation, the sample was divided into equal parts which were frozen at  $-20^{\circ}\text{C}$  and assayed during the following 25 days.

#### Interferences

Interferences were observed only in the haemolysed sera, as previously mentioned, and in the lipaemic sera. The interference in the latter sera is due to the cloudiness

Table 1. Precision studies of assay beta-glucuronidase activity.

serum value	Within-run		Between-day	
	low	high	low	high
$\bar{x}$ , IU/1	1.22	2.84	1.08	3.01
SD	.057	.035	.061	.105
CV, %	4.73	1.23	5.64	3.48
N	25	25	25	25

produced as a result of the low ratio of dilution (substrate/sample = 1:11), in which case the VP Bichromatic Analyzer gives a low energy reading.

#### Correlation with the reference method

Statistical analysis for 40 sera samples containing beta-glucuronidase from 0.6 to 5.0 IU/1 demonstrated that, in comparison with the reference method [14], the automated method performed extremely well (figure 2). The regression equation was  $y = 1.047x - 0.238$ , where  $y$  represents the results obtained with the optimized method. The correlation coefficient was 0.978.

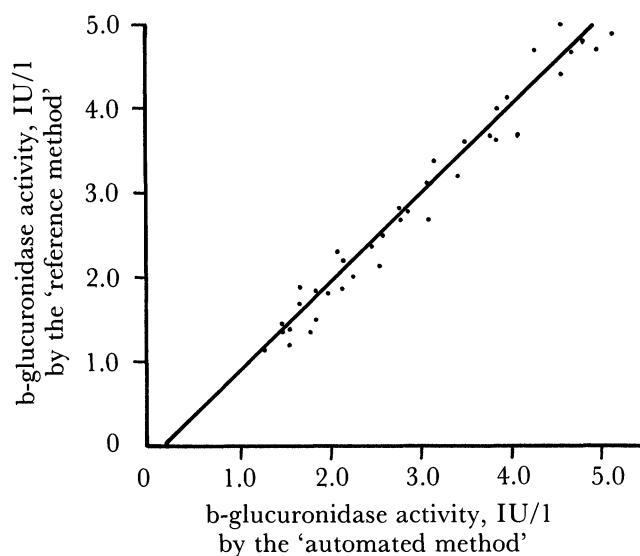


Figure 2. Comparison of the beta-glucuronidase activity, in 40 samples, determined by the 'automated method' and by the 'reference method'.

#### Reference values

Reference values were obtained from blood donors' sera. The total number was 267, of whom 207 were men and 60 women, with ages ranging from 18 to 59 years. Figure 3 shows the frequency distribution for men and women. The reference values for men range from 0.52 to 2.60 IU/1 (average 1.56 IU/1) and for women from 0.50 to 1.80 IU/1 (average 1.15 IU/1).

## Discussion

The adaptation of beta-glucuronidase analysis to the Abbott VP Bichromatic Analyzer has provided a method of analysis which is twice as fast as the reference method,

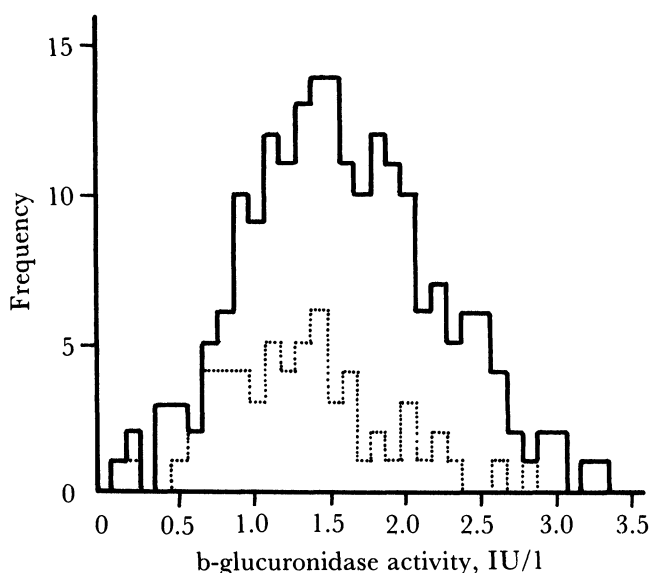


Figure 3. Distribution of beta-glucuronidase in the sera of 60 women and 207 men in a donor's population. (. . . .) female; (—) male.

and more than four times faster than the rest of the methods (the time required was reduced from 2 h to 23 min). The outstanding linearity obtained with the automated method (up to 110 IU/l) has allowed a substrate concentration of 4 mM to be used in the assay, half that of the reference method, without the risk of being outside the Michaelis-Menten linear zone as it relates to the activity values of the samples used. This 50% reduction in the substrate concentration, together with a reduction in the quantities of reagents needed, make the automated method extremely economical.

The 'within-run' and 'between-day' CVs achieved with this method for two levels of activity, normal and pathological, are better than those obtained with the optimized method used as reference. Thus, by adapting the method for determining beta-glucuronidase activity to the Abbott VP bichromatic analyzer, a method was

found that is more economical, more precise, and simpler to employ. The ease of reproduction of the results, and the close statistical correlation with the reference method, combine to create a new and useful method of beta-glucuronidase analysis.

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