

Biomimetic polymerization of acrylamide with hydrogen peroxide catalyzed by water-soluble anionic iron(III) 5,10,15,20-tetrakis-(2',6'-dichloro-3'-sulfonatophenyl)porphyrin

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Dedicated to Professor P.T. Narsimahan on his 75th birth anniversary

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Abstract

The biomimetic polymerization of acrylamide (**1**) with hydrogen peroxide catalyzed by water soluble iron(III) 5,10,15,20-tetrakis-(2',6'-dichloro-3'-sulfonatophenyl)porphyrin (**3**) gave high yield of polyacrylamide (**2**) in the molecular weight range of 1,30,000-1,50,000 g mol⁻¹ in the absence and presence of 2,4-pentanedione at an ambient temperature in nitrogen atmosphere. The molar ratio of **3**: H₂O₂: **1** was 1: 100: 3300. Yields were high at pH 4.5 but dropped at pH 7.2. Finally no polymer was formed at pH 9.2 in the absence of 2,4-pentanedione, whereas the yield raised to 99% in the presence of 2,4-pentanedione.

Keywords: Iron(III) 5,10,15,20-tetrakis-(2',6'-dichloro-3'-sulfonatophenyl)porphyrin, hydrogen peroxide, biomimetic polymerization, polyacrylamide

Introduction

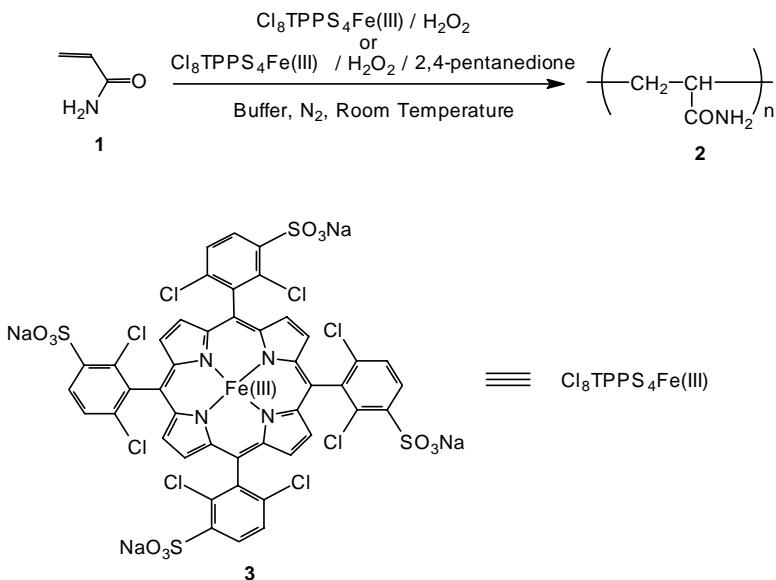
Enzymatic catalysis is involved in both in-vivo and in-vitro polymerizations.¹⁻³ Various biopolymers and non-natural synthetic polymers have been synthesized under ecofriendly and milder conditions by enzyme-catalyzed polymerizations. A selective enzyme catalyzes a specific polymerization reaction with high selectivity. Glycosidase specifically catalyses the synthesis of various natural and unnatural polysaccharides under much milder conditions with high yields.^{4,5} Lipase catalyses the synthesis of optically active polyesters and transesterification of polyesters to produce random copolyesters,^{6,7} whereas papain catalyses the polymerization of amino acids.⁸ Oxidoreductases including peroxidase, laccase and bilirubin oxidase are used as catalysts by the production of novel polyphenolics and related compounds.⁹

The reaction of phenols and anilines with hydrogen peroxide catalyzed by different oxidoreductases is an alternative ecofriendly route to synthesize the industrially important polycaromatics.⁹⁻¹⁶ Similarly the vinylic monomers have also been polymerized with hydrogen peroxide catalyzed by HRP and related enzymes under milder conditions with high yields.^{11,17-20}

Lipid-soluble and water-soluble metalloporphyrins mimic the various reactions of cytochrome P450, HRP and other monooxygenases in different reactions.^{21,22} Herein, we report a novel route for the polymerization of acrylamide with hydrogen peroxide catalyzed by anionic water-soluble iron(III) 5,10,15,20-tetrakis-(2',6'-dichloro-3'-sulfonatophenyl)porphyrin to understand the molecular mechanism of peroxidase and related oxidoreductase enzymes and their utilization in polymerization reactions.

Results and Discussion

Screening of Cl₈TPPS₄Fe(III) (3) as catalyst in polymerization of acrylamide (1). The reaction of **1** (7mmol) with H₂O₂ (0.106 mmol) in the absence of **3** gave no polymer (**Table 1, entry 1**). The reaction of **1** (7mmol) with H₂O₂ (0.106 mmol) catalyzed by **3** (2.12×10^{-3} mmol) gave polymer **2** (**Scheme 1**) in 85% yield (**Table 1, entry 3**).



Scheme 1. Polymerization of acrylamide (**1**) with hydrogen peroxide catalyzed by Cl₈TPPS₄Fe(III) (**3**).

The formation of polymer **2** was indicated in the absence of a peak at 1612 cm⁻¹ for olefinic stretching and a carbonyl absorption from 1672 to 1655 cm⁻¹ in its IR spectra. The disappearance of signals for olefinic protons at δ 5.10 and the appearance of characteristic broad signals at δ 1.59 and 2.16 corresponding to the methylene and methine protons in the ¹H-NMR spectra confirmed the formation of **2**. The average molecular weight of **2** was found to be 1,30,000 g mol⁻¹ by the viscosity measurements in water at 25°C.²³ The complete thermal degradation of **2** occurred in two stages at 315°C and 423°C when the rate of heating was 5°C/min in their thermogravimetric analysis²⁴ (**Figure 1**). The ratio of **3**: H₂O₂: **1** as 1: 100: 3300 was found to be suitable for the polymerization of **1** at room temperature.

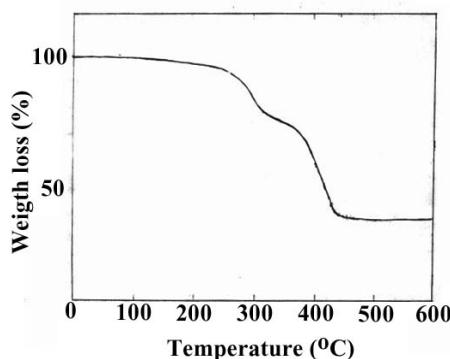


Figure 1. Thermogravimetric analysis of **2** with temperature.

Polymerization of **1 with H₂O₂ catalyzed by Cl₈TPPS₄Fe(III) (**3**) in different reaction conditions.** The reaction of **1** with H₂O₂ catalyzed by robust Cl₈TPPS₄Fe(III) (**3**) was carried out in acetate buffer (pH 4.5, 0.2M) at an ambient temperature in a nitrogen atmosphere. The monomer **1** was quantitatively consumed to give the polymer **2** in 85% yield (**Table 1, entry 3**). The yield of **2** dropped to 5% at pH 7.2 while there was no detectable amount of polymer at pH 9.2 under the same conditions (**Table 1, entry 7 and 11**).

Table 1. Effect of pH and the presence of 2,4-PD on the polymerization of **1** with H₂O₂ catalyzed by **3**

S. No.	System ^a	Yield ^b %	Mol. Wt, M _n (g mol ⁻¹) ^c	Polydispersity
1.	H ₂ O ₂ / pH 4.5	-	-	-
2.	H ₂ O ₂ / 2,4-PD ^d / pH 4.5	-	-	-
3.	3 / H ₂ O ₂ / pH 4.5	85	1,30,000	2.45
4.	3 / H ₂ O ₂ / 2,4-PD / pH 4.5	98	1,50,000	2.40
5.	H ₂ O ₂ / pH 7.2	-	-	-
6.	H ₂ O ₂ / 2,4-PD / pH 7.2	-	-	-
7.	3 / H ₂ O ₂ / pH 7.2	5	N.D.	-
8.	3 / H ₂ O ₂ / 2,4-PD / pH 7.2	15	N.D.	-
9.	H ₂ O ₂ / pH 9.2	-	-	-
10.	H ₂ O ₂ / 2,4-PD / pH 9.2	-	-	-
11.	3 / H ₂ O ₂ / pH 9.2	-	-	-
12.	3 / H ₂ O ₂ / 2,4-PD / pH 9.2	99	51,000	2.90

^a Ratio of system **3** : H₂O₂ : **1** = 1: 100 : 3300.

3: H₂O₂ : 2,4-PD : **1** = 1: 100 : 110 : 3300.

^b Yield corresponds to weight of **2** obtained to the weight of **1** used.

^c Determined by viscosity measurements at 25°C using $[\eta] = 6.8 \times 10^{-4} M_n^{0.66}$ and $[\eta] = 6.31 \times 10^{-4} M_w^{0.80}$

^d 2,4-PD = 2,4-Pentanedione, N.D.= Not determined (due to low yield).

Effect of 2,4-pentanedione (2,4-PD) on polymerization of **1 at different pH's.** The addition of 2,4-PD to the reaction of **1** with H₂O₂ catalyzed by **3** at pH 4.5 and 7.2 enhanced the yield of polymer **2** to 98 and 15% respectively (**Table 1, entry 4 & 8**). The polyacrylamide (**2**) obtained at pH 4.5 in the presence of 2,4-PD was of comparable molecular weight and polydispersity to that obtained in the absence of 2,4-PD (**Table 1, entry 3 & 4**). At pH 9.2, the addition of 2,4-PD to the above reaction facilitated the polymerization of **1** and produced the polymer **2** of low molecular weight (51,000 g mol⁻¹) and polydispersity 2.90 in 99% yield (**Table 1, entry 12**).

The polymer was obtained in 40% and 70% yield when the ratio of **3** and 2,4-PD was 1 : 5 and 1 : 50 respectively at pH 9.2. The optimal ratio of **3** and 2,4-PD was found to be 1 : 110 under given reaction conditions to obtain the polymer in 99% yield. Further increase in the concentration of 2,4-PD gave decreased product yield.

The effect of pH on the % yield of the polymer in the absence and the presence of 2,4-PD is also summarized in the **Figure 2**.

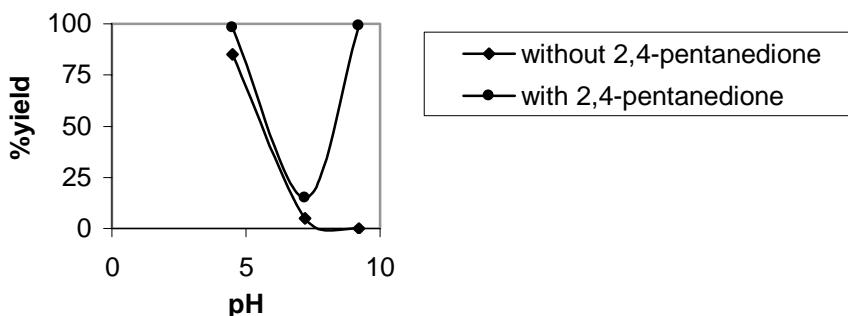


Figure 2. Effect of pH on the %yield of the polymer (**2**) in the absence and the presence of 2,4-PD.

Effect of reaction time on the molecular weight of **2.** The effect of the reaction time on the molecular weight of **2** is shown in **Figure 3**. As the reaction time increases the molecular weight of polyacrylamide increases, which indicates the free radical behaviour of the process.

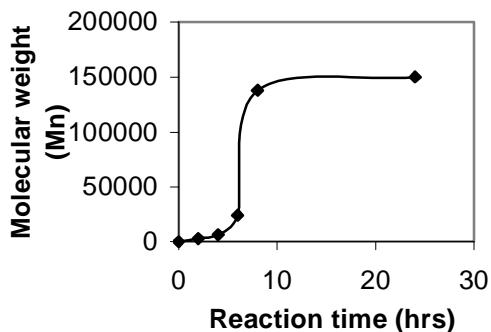
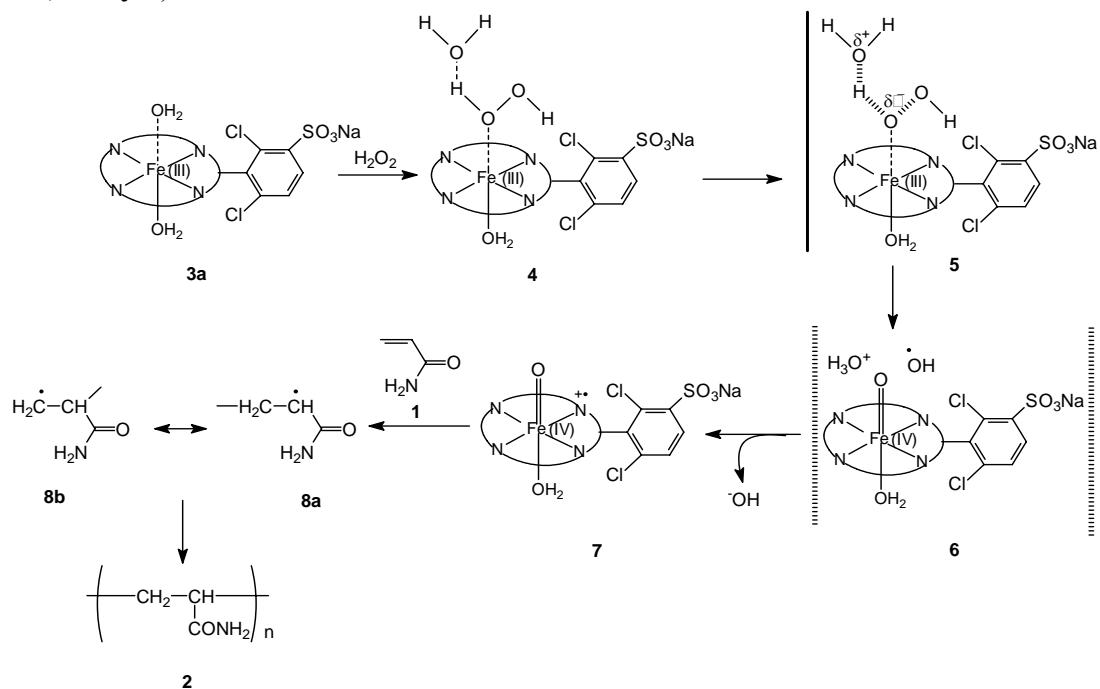


Figure 3. Changes in molecular weight of **2** produced at pH 4.5 (10 mL of 0.2M acetate buffer) using **3** : H₂O₂ : 2,4-PD : **1** in the ratio 1 : 100 : 110 : 3300 at room temperature at different reaction times.

Comparison of polymerization reaction of HRP with water-soluble Cl₈TPPS₄Fe(III) (3). The reaction of 1 with H₂O₂ catalyzed by HRP at pH 4.5 in a nitrogen atmosphere gave the polymer in 60% yield at room temperature, the concentrations being [HRP]= 2gL⁻¹, [1]= 0.46M and [H₂O₂]= 11mM. No polymer was obtained at pH 9.2 under the same conditions due to deactivation of HRP enzyme at higher pH. This kind of HRP-catalyzed polymerization of vinylic monomers has been reported using large quantities of H₂O₂ (either 1/1 or 9/1 molar ratio) with respect to monomer.^{11,19} The formation of oxoiron(IV) radical cation by the reaction of HRP and H₂O₂ has been proposed to be responsible for the polymerization reaction.

Water-soluble Cl₈TPPS₄Fe(III) (3) has been used as a model for HRP to mimic their various reactions. At acidic pH, iron(III) porphyrin exists as -Fe(III)(H₂O)₂- (3a).^{25,26} On the addition of H₂O₂ to 3a, the species -Fe(III)(OOH)(H₂O)- (4) is generated. 4 then spontaneously gives the highvalent oxoiron(IV) radical cation (7) through the transient intermediates 5 and 6. The intermediate 7 is responsible for initiating the formation of acrylamide radical (8a and 8b), which attacks the other acrylamide molecule to form a bigger free radical and hence the reaction further propagates to give the polymer 2. The reaction is terminated either by the combination of two bigger radicals or by abstraction of a proton from the solvent or another monomer molecule (**Scheme 2**). The stable dimethoxyiron(IV) porphyrin is generated by reaction of iron(III)porphyrin with H₂O₂ in presence of large excess of methanol.²⁷ Thus the polymerization of 1 is quenched by the addition of large excess of methanol. The formation of intermediate 7 could be explained only by the heterolytic cleavage of H₂O₂, which leads to the polymerization. This was confirmed as no polymer was obtained in the presence of H₂O₂ and absence of 3 (**Table 1, entry 1**).



Scheme 2. Proposed mechanism for the formation of reactive intermediate in polymerization of 1 with hydrogen peroxide catalyzed by Cl₈TPPS₄Fe(III) (3).

At neutral and near alkaline pH, the peroxide anion ($\text{^{\cdot}OOH}$) from H_2O_2 is not able to replace the axial ligand OH on $-\text{Fe(III)}(\text{H}_2\text{O})(\text{OH})-$, which is the species when **3** is present in the buffer of neutral and higher pH. Thus the concentration of highvalent oxoiron(IV) radical cation (**7**) decreases and hence very low or no polymerization is observed at these pH's.

The polymerization of vinylic monomers with H_2O_2 catalyzed by HRP in the presence of 2,4-PD and other β -diketones increases the yield of polymer significantly.^{17-18,20} Polyacrylamide of molecular weight 1,24,000 g mol⁻¹ and polydispersity 2.5 has been obtained by the polymerization of **1** with hydrogen peroxide catalyzed by HRP in the presence of 2,4-PD using H_2O_2 -to-monomer ratio of 1/66 mol/mol at pH 5.1 while no polymer is reported in the absence of 2,4-PD.¹⁷ Thus 2,4-PD was added to the reaction of **1** with H_2O_2 catalyzed by $\text{Cl}_8\text{TPPS}_4\text{Fe(III)}$ (**3**) at pH 4.5 and 7.2 and it enhanced the yield of polymer **2** to 98 and 15% respectively. At pH 4.5, the polymer **2** of comparable molecular weight (1,30,000 g mol⁻¹) and polydispersity (2.45) was obtained when **3** was used as the catalyst in place of HRP without any need for the addition of 2,4-PD. The addition of 2,4-PD at pH 9.2 offered a new route for polymerization of **1** at this pH, where the use of HRP is limited owing to its deactivation. The reaction of **1** with H_2O_2 in the presence of 2,4-PD in the absence of **3** produced no polymer (**Table 1, entry 10**). At pH 9.2, the reaction of **3** with H_2O_2 in the presence of 2,4-PD may produce a stable intermediate which contributes to the facile polymerization of **1** at this pH.

The large excess of H_2O_2 with respect to the monomer resulted in the degradation of the iron(III) porphyrin. Thus, a 1:66 molar ratio of H_2O_2 to monomer was found to be suitable for the polymerization of **1** both in the absence and the presence of 2,4-PD.

Conclusions

The water soluble anionic $\text{Cl}_8\text{TPPS}_4\text{Fe(III)}$ (**3**) in the presence of hydrogen peroxide is an economical and suitable system to initiate the polymerization of acrylamide at acidic and neutral pH, by using the molar ratio of **3**: H_2O_2 : **1** as 1 : 100 : 3300. The addition of 2,4-PD in this system enhances the yield of the polymer at both acidic and neutral pH and also facilitates the polymerization of **1** at alkaline pH, where HRP is deactivated. Thus the water-soluble anionic iron(III) porphyrin and H_2O_2 mimic the polymerization reaction of oxidoreductase enzyme HRP in the presence or absence of 2,4-PD under milder conditions with excellent yield.

Experimental Section

Material and methods. **1** and 2,4-PD were obtained from Biochemicals unit, India and Merck, Munchen, respectively and were also purified before use. The aqueous hydrogen peroxide (30% v/v) was obtained from CDH India and used without further purification. The 5,10,15,20-tetrakis-(2',6'-dichloro-3'-sulfonatophenyl)porphyrin ($\text{H}_2\text{Cl}_8\text{TPPS}_4$) was prepared by

sulphonation of 5,10,15,20-tetrakis-(2',6'-dichlorophenyl)porphyrin (Cl_8TPPH_2) with oleum at 130°C by following the literature procedure and metallated with ferrous chloride in DMF.^{28,29}

The average molecular weight of polyacrylamide was determined from intrinsic viscosity measurements according to Mark-Houwink equations.²³ Viscosity measurements were obtained at 25°C in water using ubbelohde viscometer and $[\eta]_{c \rightarrow 0}$ was evaluated by the extrapolation of experimental data. $[\eta]_{c \rightarrow 0}$ values were used to calculate the number and weight average molecular weight according to the relations $[\eta] = 6.8 \times 10^{-4} M_n^{0.66}$ and $[\eta] = 6.31 \times 10^{-4} M_w^{0.80}$. ¹H NMR spectra were recorded on a Bruker Avance Spectrospin 300MHz spectrophotometer. Perkin Elmer FT-IR Spectrum 2000 Spectrometer was used to record IR spectra. Thermogravimetric analysis was obtained on a Rigaku Thermoflex, PTC-10A.

Polymerization of 1 with hydrogen peroxide catalyzed by water-soluble $\text{Cl}_8\text{TPPS}_4\text{Fe(III)}$ (3). The solution of **1** (7 mmol in 10mL acetate buffer of pH 4.5) was charged into a three-necked round-bottomed flask and degassed for 15 minutes. $\text{Cl}_8\text{TPPS}_4\text{Fe(III)}$ (2.12×10^{-3} mmol in 0.5 mL buffer) and 30% H_2O_2 (0.106 mmol) were successively injected into the above flask under stirring. The reaction was stirred for a predetermined time at an ambient temperature under a nitrogen atmosphere. The polymer was obtained by quenching the reaction with a large excess of methanol and filtered off, washing with methanol and drying under vacuum at 50°C. The yield and molecular weight of polymers are given in **Table 1**.

Polymerization of 1 with hydrogen peroxide catalyzed by water-soluble $\text{Cl}_8\text{TPPS}_4\text{Fe(III)}$ (3) in the presence of 2,4-PD. The procedure and molar ratio of reactant (**1**) and catalyst (**3**) used in this case were exactly the same as mentioned above except for the addition of 2,4-PD (0.18 mmol) which was added simultaneously along with hydrogen peroxide. The results obtained in this case are also summarized in **Table 1**.

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References

1. Kobayashi, S.; Uyama, H.; Kimura, S. *Chem. Rev.* **2001**, *101*, 3793.
2. Gross, R. A.; Kumar, A.; Kalra, B. *Chem. Rev.* **2001**, *101*, 1097.
3. Uyama, H.; Kobayashi, S. *J. Mol. Cat. B: Enzymatic* **2002**, *19*, 117.
4. Kobayashi, S.; Kashiwa, K.; Kawasaki, T.; Shoda, S. *J. Am. Chem. Soc.* **1991**, *113*, 3079.
5. Kobayashi, S.; Kiyosada, T.; Shoda, S. *J. Am. Chem. Soc.* **1996**, *118*, 13113.
6. Runge, M'; O'Hagan, D.; Haufe, G. *J. Polym. Sci., Polym. Chem. Ed.* **2000**, *38*, 2004.

7. Kumar, A.; Kalra, B.; Dekhterman, A.; Gross, R. A. *Macromolecules* **2000**, *33*, 6303.
8. Anderson, G.; Luisi, P. L. *Helv. Chim. Acta* **1979**, *62*, 488.
9. Ikeda, R.; Sugihara, J.; Uyama, H.; Kobayashi, S. *Macromolecules* **1996**, *29*, 8702.
10. Uyama, H.; Kurioka, H.; Sugihara, J.; Komatsu, I.; Kobayashi, S. *Bull. Chem. Soc. Jpn.* **1995**, *68*, 3209.
11. Uyama, H.; Loavisavapanich, C.; Ikeda, R.; Kobayashi, S. *Macromolecules* **1998**, *31*, 554.
12. Dordick, J. S.; Marletta, M. A.; Klibanov, A. V. *Biotech. Boeing*. **1987**, *30*, 31.
13. Uyama, H.; Kurioka, H.; Kaneko, I.; Kobayashi, S. *Chem. Lett.* **1994**, 423.
14. Akkara, J. A.; Senecal, K. J.; Kaplan, D. L. *J. Pol. Sci. Part A* **1991**, *29*, 1561.
15. Premchandran, R. S.; Banerjee, S.; Wu, X. K.; John, V. T.; McPherson, G. L. *Macromolecules* **1996**, *29*, 6452.
16. Kobayashi, S.; Kaneko, I.; Uyama, H. *Chem. Lett.* **1992**, 393.
17. Teixera, D.; Lalot, T.; Brigodiot, M.; Marechal, E. *Macromolecules* **1999**, *32*, 70.
18. Kalra, B. Gross, R. A. *Biomacromolecules* **2000**, *1*, 501.
19. Derango, A. R.; Chiang, L.-C.; Dowbenko, R.; Lasch, J. G. *Biotech. Tech.* **1992**, *6*, 523.
20. Singh, A.; Ma, D.; Kaplan, D. L. *Biomacromolecules* **2000**, *1*, 592.
21. Chauhan, S. M. S.; Kalra, B.; Mohapatra, P. P. *J. Mol. Cat. A: Chemical* **1999**, *137*, 85.
22. Chauhan, S. M. S.; Sahoo, B. B. *Bioorg. Med. Chem.* **1999**, *7*, 2629.
23. Shawki, S. M.; Hamielec, A. E. *J. Appl. Polym. Sci.* **1979**, *23*, 3323.
24. Yang, M. H. *Polymer Testing* **1998**, *17*, 191.
25. Panicucci, R.; Bruice, T. C. *J. Am. Chem. Soc.* **1990**, *112*, 6063.
26. Bruice, T. C. *Acc. Chem. Res.* **1991**, *24*, 243.
27. Meunier, B. *Chem. Rev.* **1992**, *92*, 1411.
28. Dolphin, D.; Nakano, T.; Maioni, T. E.; Kirk, T. K.; Farrell, R. In *Lignin Enzymic and Microbial Degradation*; Odier, E. Ed.; INRA: Paris, 1987, 157.
29. Chauhan, S. M. S.; Ray, P. C.; Satapathy, S.; Vijayraghavan, B. *Ind. J. Chem.* **1992**, *31B*, 837.