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Numerical Characterization of DNA Primary Sequence

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Numerical Characterization of DNA Primary Sequence[#]

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Abstract

In a previous paper, the authors defined a numerical characterization of DNA primary sequences. A DNA primary sequence was reduced to a few of binary sequences, based on the classifications of the four nucleic acid bases. The reduced sequences are called the characteristic sequences. For each characteristic sequence, we associated two 2×2 matrices, the elements of which are given by the frequency of occurrence of all (0,1) triplets in the characteristic sequence. In this paper, we use eigenvalues of the new matrices to characterize the biological functions of purine–pyrimidine, amino–keto groups and weak–strong H–bonds, respectively.

Keywords. DNA primary sequence; characteristic sequences; leading eigenvalue; similarity; dissimilarity.

1 INTRODUCTION

With the imminent completion of the Human Genome Project and the fast increase of many complete genomes of prokaryote and eukuaryote, fundamental questions regarding the characteristics of these sequences arise, the first of which is how to compare genomes. Hence analysis and understanding of the DNA primary sequences are very important tasks in bioinformatics.

Usually, a DNA primary sequence can be taken as a string of letters A, G, C, T, which denote the four nucleic acid bases: adenine, guanine, cytosine and thymine, respectively. Therefore, the analysis and understanding of DNA primary sequences are performed via comparisons of such strings of the four letters. In previous research, the comparisons of DNA primary sequences are mainly to consider the alignment of the DNA primary sequences. The alignment of sequences is performed by the computer to find the smallest number of changes (deletions, insertions, substitutions, shifts) that are necessary to match labels in two DNA primary sequences.

[#] Dedicated to Professor Haruo Hosoya on the occasion of the 65th birthday.

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Some researchers consider graphical representations for DNA primary sequences [1–17], in particular, Gates [1], Hamori and Ruskin [2], Leong and Morgenthaler [4], Randić [9–13], Nandy [5–8,13–15], Zhang [16,17] and others, considering a real DNA primary sequence as a curve embedded in 2–D plane or 3–D space. Using research on the graphical representations, we can derive some numerical characterization for DNA primary sequences.

An alternative approach of the comparisons for DNA sequences is suggested by Randić *et al.* [9–13], who considered mathematical invariants of DNA primary sequences rather than the sequences themselves. For chemical structure and chemical graphs we can in fact obtain numerous invariants that are applied for characterization and comparison of structures. There are hundreds of topological indices that have been used in structure–property–activity studies based on molecular graphs. In this way, we can arrive at invariants for DNA primary sequences to associate a matrix with a DNA primary sequence. Once a matrix representation of sequences is given, one can consider suitable matrix invariants as invariants of the comparison of DNA sequences.

In a previous paper [3], we introduced another representation for DNA sequences, which is based on the idea of the coarse–grained description of the DNA primary sequence: we classify the four nucleic acid bases into two groups, purine–pyrimidine, amino–keto groups and weak–strong H–bonds, respectively, and then label the bases of purine, amino and weak H–bonds by 1, and the bases of pyrimidine, keto and strong H–bonds by 0, respectively. Thus, from a DNA primary sequence we obtained three (0,1)–sequences, which are called the characteristic sequences of the DNA primary sequence. For each characteristic sequence we constructed a set of 2×2 matrices, which are based on counting of the frequency of occurrence of all (0,1) triplets of the characteristic sequence. The leading eigenvalues of these matrices are computed and considered as invariants for the comparison of DNA primary sequences.

Species	Sequence	Length				
Human	ATGGTGCACCTGACTCCTGAGGAGAAGTCTGCCGTTACTGCCCTGTG	02				
	GGGCAAGGTGAACGTGGAGTAAGTTGGTGGTGAGGCCCTGGGCAG					
Coat	ATGCTGACTGCTGAGGAGAAGGCTGCCGTCACCGGCTTCTGGGG	97				
Goat	CAAGGTGAAAGTGGATGAAGTTGGTGCTGAGGCCCTGGGCAG	80				
Gallug	ATGGTGCACTGGACTGCTGAGGAGAAGCAGCTCATCACCGGCCTCTG	02				
Gallus	GGGCAAGGTCAATGTGGCCGAATGTGGGGCCGAAGCCCTGGCCAG	92				
Onegaum	ATGGTGCACTTGACTTCTGAGGAGAAGAACTGCATCACTACCATCTG					
Opossum	GTCTAAGGTGCAGGTTGACCAGACTGGTGGTGAGGCCCTTGGCAG					
Lomur	ATGACTTTGCTGAGTGCTGAGGAGAATGCTCATGTCACCTCTCTGTG					
Lemu	GGGCAAGGTGGATGTAGAGAAAGTTGGTGGCGAGGCCTTGGGCAG	92				
Mouso	ATGGTGCACCTGACTGATGCTGAGAAGGCTGCCGTTACTGCCCTGTG					
Mouse	GGGCAAGGTGAACGTGGATGAAGTTGGTGGTGAGGCCCTGGGCAGG					
Rabbit	ATGGTGCATCTGTCCAGTGAGGAGAAGTCTGCGGTCACTGCCCTGT	00				
	GGGGCAAGGTGAATGTGGAAGAAGTTGGTGGTGAGGCCCTGGGC					
Rat	ATGGTGCACCTAACTGATGCTGAGAAGGCTACTGTTAGTGGCCTGTG GGGAAAGGTGAACCCTGATAATGTTGGCGCTGAGGCCCTGGGCAG	92				

In this paper, through comparison of characteristic sequences we try to find the biological functions of purine–pyrimidine, amino–keto groups and weak–strong H–bonds, respectively. In Table 1, the exon–1 of the β –globin gene for eight species are listed, which were reported by Randić [9].

2 CONSTRUCTION OF THE CHARACTERISTIC SEQUENCES

Nucleic acids and proteins are all linear macromolecules. However, comparison of DNA primary sequences should be considered not only the string structures, but also their chemical structures. In DNA primary sequences, the four bases A, C, G, T can be divided into two classes according to their chemical structures: purine $R = \{A, G\}$ and pyrimidine $Y = \{C, T\}$, or amino group $M = \{A, C\}$ and keto group $K = \{G, T\}$. Besides these, the division can be also made according to the strength of the hydrogen bond, *i.e.*, weak H–bonds $W = \{A, T\}$ and strong H–bonds $S = \{G, C\}$.

Let $S = a_1 a_2 a_3 \cdots$ be a DNA primary sequence. Using above classifications, we can transform a DNA primary sequence into three (0,1) sequences by three homomorphic maps ϕ_i , i = 1, 2, 3, $\phi_i(S) = \phi_i(a_1)\phi_i(a_2)\cdots$, as follows:

$$\phi_1 = \begin{cases} 1 & \text{if} \quad a_i \in R \\ 0 & \text{if} \quad a_i \in Y \end{cases}$$

$$\phi_2 = \begin{cases} 1 & \text{if} \quad a_i \in M \\ 0 & \text{if} \quad a_i \in K \end{cases}$$

$$\phi_3 = \begin{cases} 1 & \text{if} \quad a_i \in W \\ 0 & \text{if} \quad a_i \in S \end{cases}$$

Thus, we obtain three (0,1) sequences corresponding to the same DNA primary sequence, and we call them as (R,Y)-, (M,K)- and (W,S)-characteristic sequences of the DNA primary sequence, respectively.

In [3], we constructed a set of 2×2 matrix and computed their leading eigenvalues for three characteristic sequences. Using the leading eigenvalues, we compared the similarities and dissimilarities for eight species in Table 1. The results in [3] coincide with the result of Randić's papers. It demonstrates that the comparison of three characteristic sequences is the same as the comparison of DNA primary sequences.

As we have seen in [3], the three characteristic sequences contain all information of the primary sequence. On the other hand, each characteristic sequence is a coarse–grained description for the DNA primary sequence, *i.e.*, some information for DNA primary sequence may be lost in a characteristic sequence so that different DNA primary sequences may have certain similar characteristic sequences. This just reflects the functions of the classifications. Therefore, comparing

each characteristic sequences has special significance. In Table 2, we list the characteristic sequences of the eight DNA sequences of Table 1. In the next section we will compare each characteristic sequence and get some conclusions that cannot be obtained from direct comparison of DNA primary sequences.

Table 2. Characteristic Sequences of the Eight DNA Sequence from Table

human	
101101010001100000111111111100010010010	
10000011110011011001001011001001100011001110000	
1100100100101010100101010101010000111010	
goat	-
101001100100111111111100100100100100110000	
100100110010010010110010011001111001001	
11001010100101010010100001010000011010000	
gallus	
1100100101001010010101010100101101000000	
onossim	-
lamur	-
	_
rabbit	
101101010001000110111111111100010110010010000	
10000011010001110001001011001001000111001110000	
11001001101001010101010101010100001010000	
rat	
10110101000110011010011111110010010011011000101	
1000001111011100100100101011001011000010000	
1100100100111010110010101100011010111010	

3 COMPARISON OF CHARACTERISTIC SEQUENCES

In [3], we also introduced a 2×2×2 cubic matrix with 8 entries $f_{ijk}^{X} = (100m_{ijk}^{X})/(N-2)$, where m_{ijk}^{X} is the enumeration of the (0,1) triplet *ijk* in characteristic sequence X and N is the length of X. Clearly, it represents the 100 times of the frequency of occurrence of the (0,1) triplet *ijk* in X. That we take the 100 times is for convenience of tabulation and computation. By F^{R} , F^{M} and F^{W} we denote the cubic matrices for the (*R*,*Y*)–, (*M*,*K*)– and (*W*,*S*)–characteristic sequences, respectively. We partition each of the cubic matrices into a pair of 2×2 condensed matrices F_{0}^{X} and F_{1}^{X} , where

 $F_0^X = (f_{0jk}^X)$ and $F_1^X = (f_{1jk}^X)$ with *X* being *R*, *M* or *W*. In [3], we computed the leading eigenvalues condensed matrices as above. In Table 3, all leading eigenvalues of characteristic sequences are listed, as reported in [3].

Table 3. Leading eigenvalues of the 6 matrices F_0^X and F_1^X for the eight DNA sequences of Table 1.

			*	-		
Species	F_0^R	F_1^R	F_0^M	F_1^M	F_0^W	F_1^w
Human	21.7	28.9	31.3	18.3	30.0	22.2
Goat	20.3	30.8	30.2	21.7	30.4	21.4
Gallus	22.6	28.2	26.5	23.2	33.2	20.7
Opossum	23.0	26.6	27.6	21.8	26.6	25.0
Lemur	21.1	30.3	33.2	20.4	26.5	22.9
Mouse	21.6	29.6	31.5	19.9	29.6	23.4
Rabbit	20.5	31.7	34.7	18.6	29.2	22.2
Rat	22.8	28.3	30.3	21.3	28.2	22.3

For each characteristic sequence, we take the leading eigenvalue as a two-dimensional vector (F_0^x, F_1^x) , by which we compare the (R, Y)-, (M, K)- and (W, S)-characteristic sequences of DNA primary sequences based on the Euclidean distance between the end points of the two-dimensional vectors, respectively. The results of comparisons are listed on the three tables, where Table 4 reveals the information of purine-pyrimidine group, Table 5 the information of amino-keto group, and Table 6 the information of weak-strong H-bonds, respectively.

Table 4. Similarity/dissimilarity table for the eight DNA sequences of Table 1 based on their (R, Y) characteristic sequences.

Species	Goat	Gallus	Opossum	Lemmur	Mouse	Rabbit	Rat
Human	2.36008	1.14018	2.64197	1.52315	2.86007	3.04631	1.25300
Goat		3.47131	4.99300	0.94339	5.22015	0.92195	3.53553
Gallus			1.64924	2.58070	1.78885	4.08167	0.22361
Opossum				4.15933	0.40000	5.67979	1.71172
Lemmur					4.3566	1.52315	2.62488
Mouse						5.86686	1.80278
Rabbit							4.10488

Table 5. Similarity/dissimilarity table for the eight DNA sequences of Table 1 based on their (M,K) characteristic sequences.

Species	Goat	Gallus	Opossum	Lemmur	Mouse	Rabbit	Rat
Human	3.57351	6.8593	5.02096	2.83196	1.71172	3.41321	3.16228
Goat		3.99249	2.50200	3.26956	2.14009	5.46443	0.41231
Gallus			1.84391	7.26154	5.93633	9.40213	4.24853
Opossum				5.67539	4.20476	7.69675	2.64764
Lemmur					1.74642	2.34307	3.03645
Mouse						3.49285	1.76918
Rabbit							5.16236

Observing Tables 4, 5 and 6, we can obtain some information for each characteristic sequence. For example, the species gallus is the most dissimilarly with others in Table 6. However, we do not see the same result from Table 4 and 5, even the value of gallus–rat pair is the least in Table 4. The

species gallus is the only non-mammalian species among these considered species. Whether or not this means that the essential nature of the mammalian species may be revealed mainly in the characteristic sequence of weak-strong H-bonds group. On the other hand, the results in Tables 5 and 6 are very similar to that of the comparison of DNA primary sequences. This means that the information of the similarities for eight sequences may contain mainly in the reduce sequences of amino-keto groups and weak-strong H-bonds groups.

Table 6. Similarity/dissimilarity table for the eight DNA sequences of Table 1 based on their (W,S) characteristic sequences.

Species	Goat	Gallus	Opossum	Lemmur	Mouse	Rabbit	Rat
Human	0.894427	3.53412	4.40454	3.56931	1.96977	0.80000	1.80278
Goat		2.88617	5.23450	4.17852	2.72029	1.44222	2.37697
Gallus			7.87718	7.05195	5.50364	4.2720	5.24976
Opossum				2.10238	2.56125	3.82099	3.13847
Lemmur					1.70294	2.78927	1.80278
Mouse						1.28062	0.70000
Rabbit							1.00499

Furthermore, we can observe the least value in each table: the gallus-rat pair in Table 4, the goat-rat pair in Table 5, and the mouse-rat pair in Table 6, respectively. Whether these results imply that the three characteristic sequences reflect some intrinsic essence of species rat from different aspect.

Generally, we can also observe the least value of all species in each table, so that we can obtain information of the (R,Y)-, (M,K)- and (W,S)-characteristic sequences, respectively. For example, the mouse species, the least value in Tables 4–6 are the opossum, human and rat, respectively. These results illuminate that the three characteristic sequences reflect some essence of mouse species from different aspect.

4 CONCLUSIONS

Comparing characteristic sequences, we can get some information that cannot be obtained from the direct comparison of DNA primary sequences and observe some special nature in species from different aspect. Although some information may be lost in characteristic sequences, we can focus our attention on the information of our interest. This is the advantage of our approach.

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